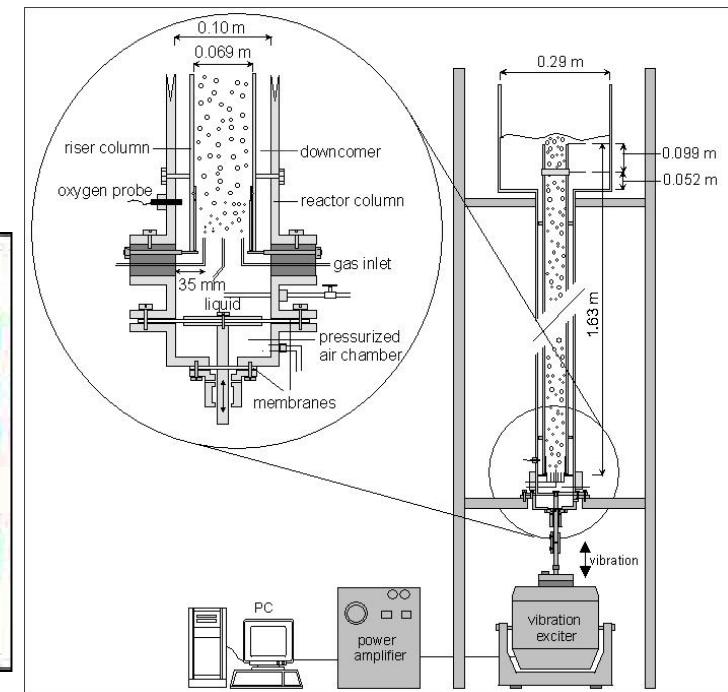
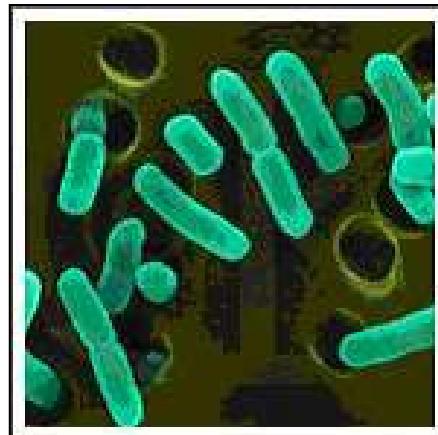


# Einführung in die Bioverfahrenstechnik 1

## Grundlagen der Bioverfahrenstechnik



Prof. Dr. Axel Blokesch - erreichbar: [blokesch@fb2.fra-uas.de](mailto:blokesch@fb2.fra-uas.de)

Büro: Gebäude 8, Raum 212 a Tel. z.Z. nicht angeschlossen.

Als Prüfungsausschussvorsitzender auch Ansprechpartner für die Anerkennung von Leistungen aus einem vorhergehenden Studium und Planung eines Auslandsaufenthaltes

**Vorlesungen:** Donnerstag, den 20.11. und 27.11.2024  
von 10:00 bis 11:30 in HC 113 (Hungener Str.)

**Exkursion:** Montag, den 16.12.2024

Treffpunkt Tor Ost des Industrieparks Höchst um 11:15

Besuch der Lantus-Produktionsanlage (Langzeit-Insulinanalogen)  
um 12:30

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## Tasks of the process development:

- choice of a reactor type
- optimization of the process
- scale-up

## Possible demands of bioreactors:

- sufficient residence time
- high oxygen transfer (in case of aerobic fermentation), good carbon dioxide transfer and removal of excess oxygen in case of oxygenic photosynthetic organisms, e.g. green algae, cyanobacteria
- in the case of photosynthetic organisms also efficient illumination
- no sedimentation of biomass
- good mix of the compounds
- no accumulation of inhibitors
- small mechanical stress
- compliance with temperature, pH
- prevention of foam formation (→ *mechan.*: foam breaker; *chemic.*: anti-foam)
- Sterilization and simple cleaning (→ “Sterilization in Place”, “Cleaning in Place”)
- flexible application

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# Basic Differences between Bioreactors

## 1. Distribution of cells

surface culture, submerged culture

## 2. Different forms of reactors

stirred tank ~, bubble column ~, air lift reactor

## 3. Operational mode

batch, fed-batch, continuous

## 4. Energy supply

nutrient media,

*in case of photosynthetic cellular systems: irradiation with light*

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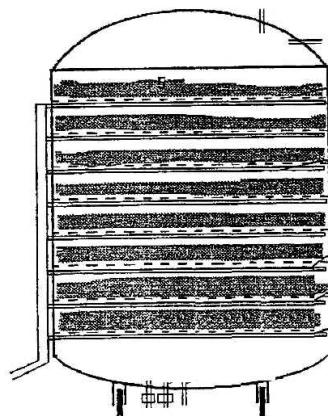
# Basic Differences between Bioreactors

## 1. Distribution of cells

### surface culture

- Substrate on trays
- Inoculation with microorganisms (preferably on substrate surface)
- Realization preferably under aerobe conditions

Prophyta, US Patent 6620614



### submerged culture

- Equal distribution of micro-organisms within the substrate by stirring or flow
- Oxygen supply (preferably from the bottom of the culture medium)
- Realization either under anaerobe and aerobe conditions

# Growing Microorganisms

**Surface Cultivation**  
on solid nutrients

**Submerged Fermentation**  
in liquid culture

on a lab scale:

- agar plates
- slant agar tubes

- shaker flasks

large scale production

• Solid Surface Fermentation  
(SSF)

- Stirred Tank Reactor
- Air Lift Reactor

# Growing Microorganisms

**Surface Cultivation**  
on solid nutrients

**Submerged Fermentation**  
in liquid culture

on a lab scale:

- agar plates



- slant agar tubes

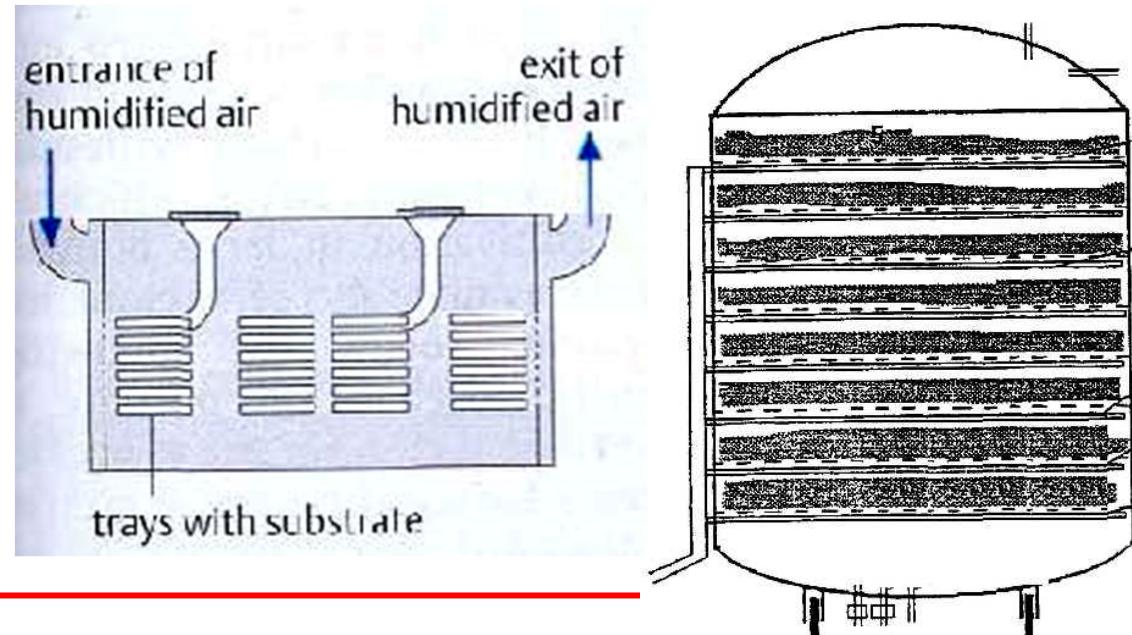
- shaker flasks



ments in a c  
ce

**G25 New Brunswick Floor Model Incubator**

## Surface Bioreactor



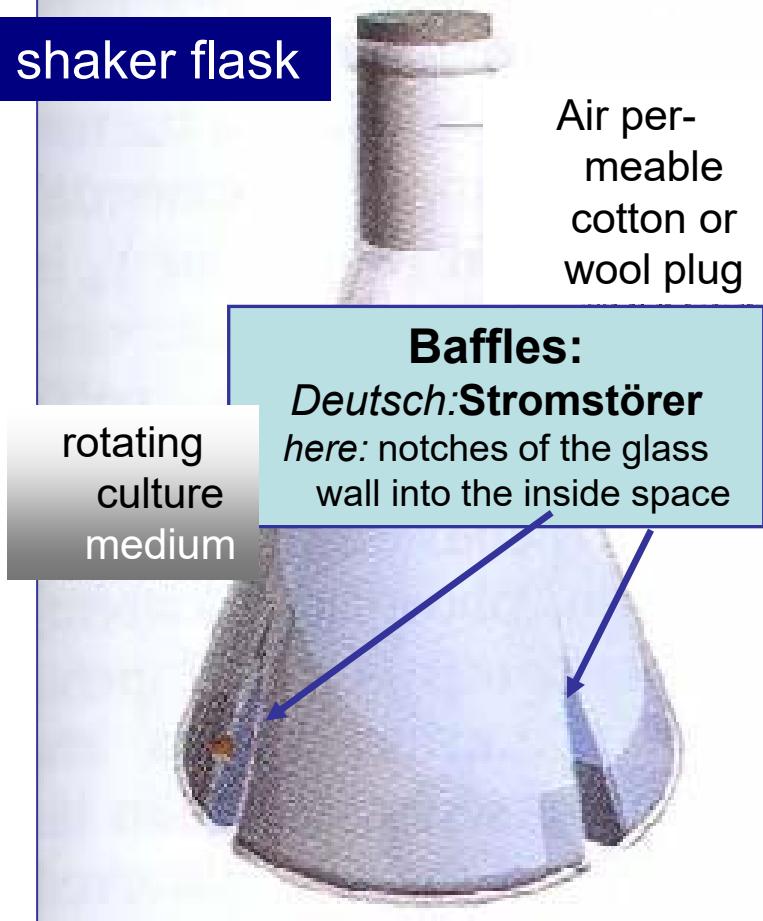
Prophyta, US Patent 6620614

- + simple to operate
- + space-time-yield rather low
- often unequal distribution of microorganisms
- applications:
  - production of citric acid with *Aspergillus niger* - *previously*
  - aerobic waste water treatment - *frequently*
  - enzyme production – *occasionally, frequently in SE Asia*

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## Different forms of bioreactors for submerged fermentation

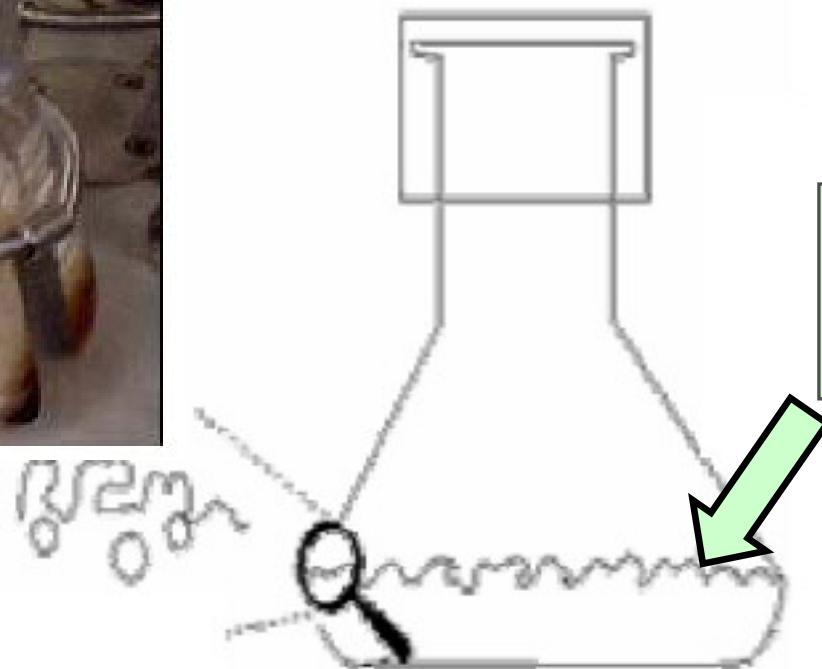
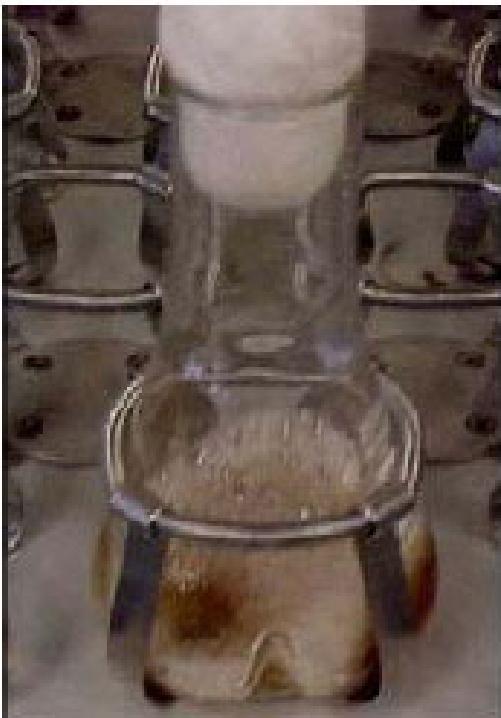
shaker flask



stirred tank reactor



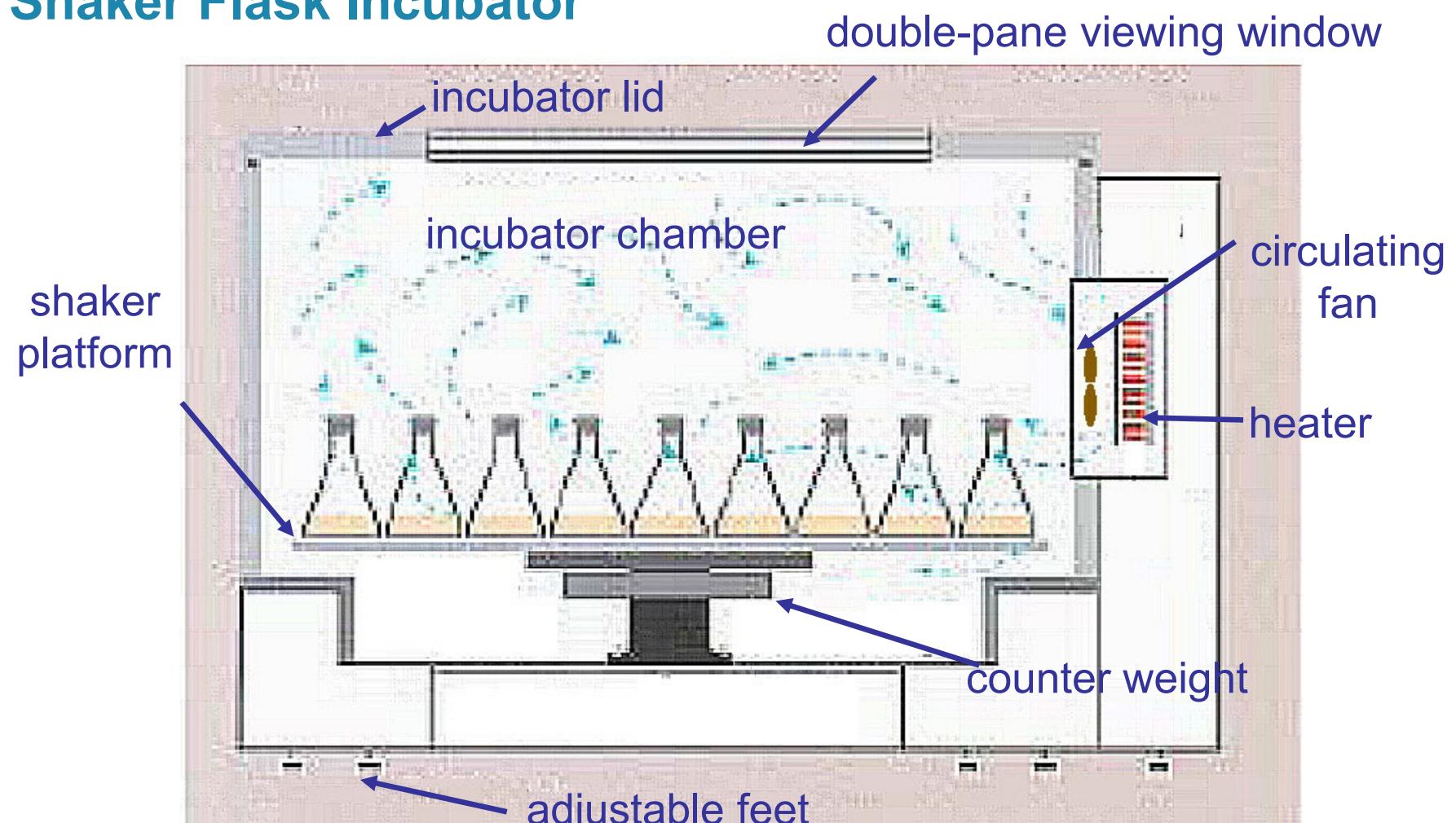
# Shaker Flask Incubator



Agitation increases  
the surface area for  
oxygen transfer

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# Shaker Flask Incubator



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# Schüttelkultur-Inkubator



**Modell von New Brunswick, bis März 2024 im Bio-Labor in Gebäude 7, 2. OG  
Bis 2023 wurde im Rahmen von Projekten und Abschlussarbeiten die Grünalge *Chlorella vulgaris* kultiviert.**

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# Submerged Fermentation

**Stirred Tank Reactor**



Braun Fermenter at DSM Biopract GmbH

aeration by mechanical agitation

**Bubble Column Reactor**

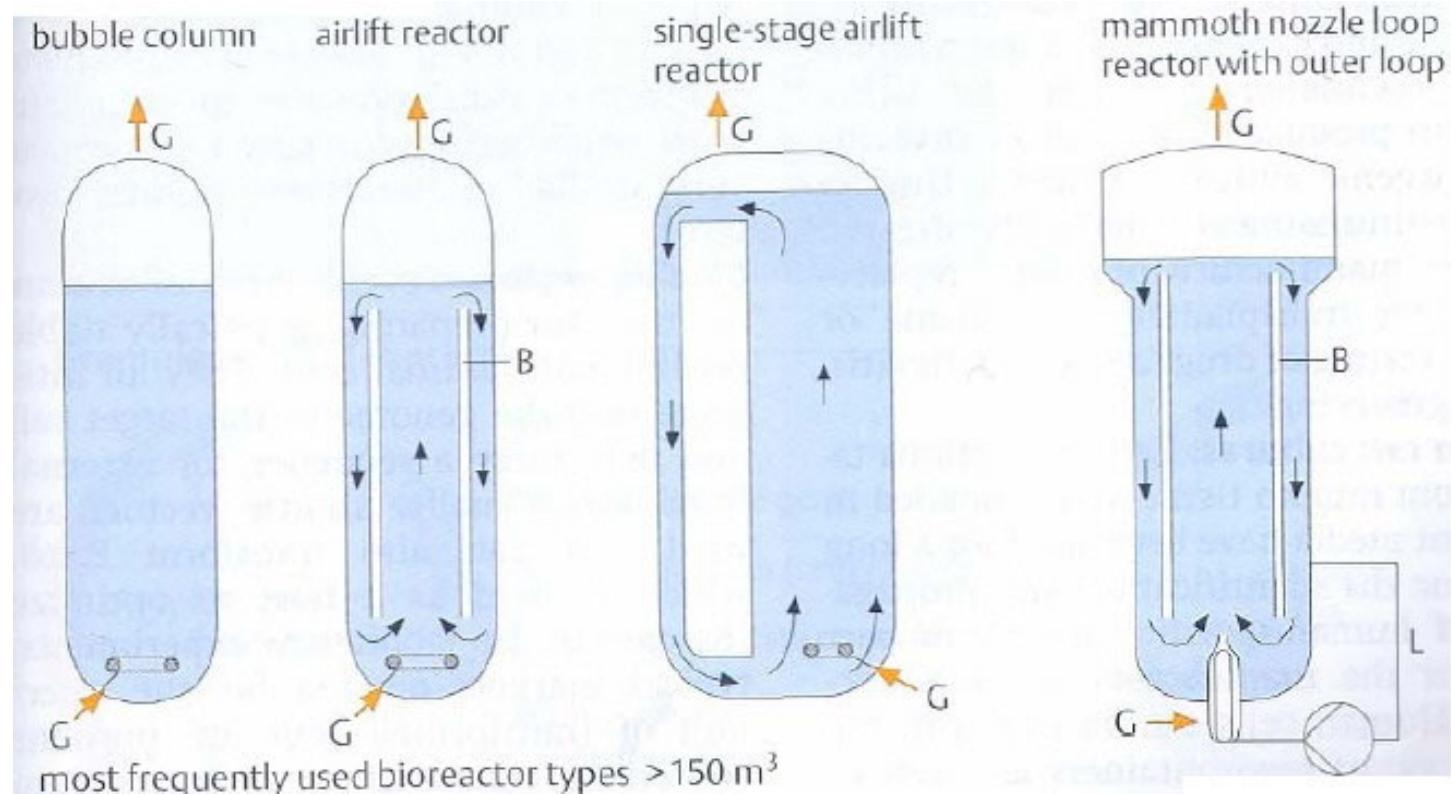


bubbling air into media for aeration  
and for mixing of the reaction media

# Submerged Fermentation

**Bubble Column Reactor**  
*Deutsch: Blasensäule*

**Air Lift Reactors**



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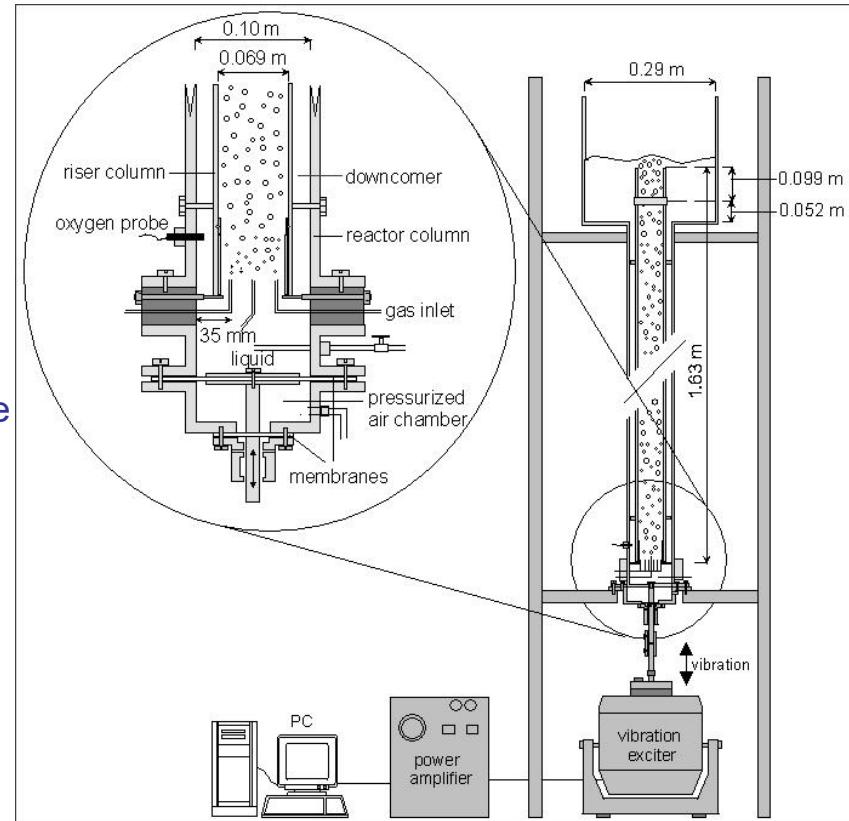
# Submerged Fermentation

## Air Lift Reactor

**Aeration from the bottom →  
Air cycles back from the top to  
the bottom through an  
internal or external loop.**

**also possible:**

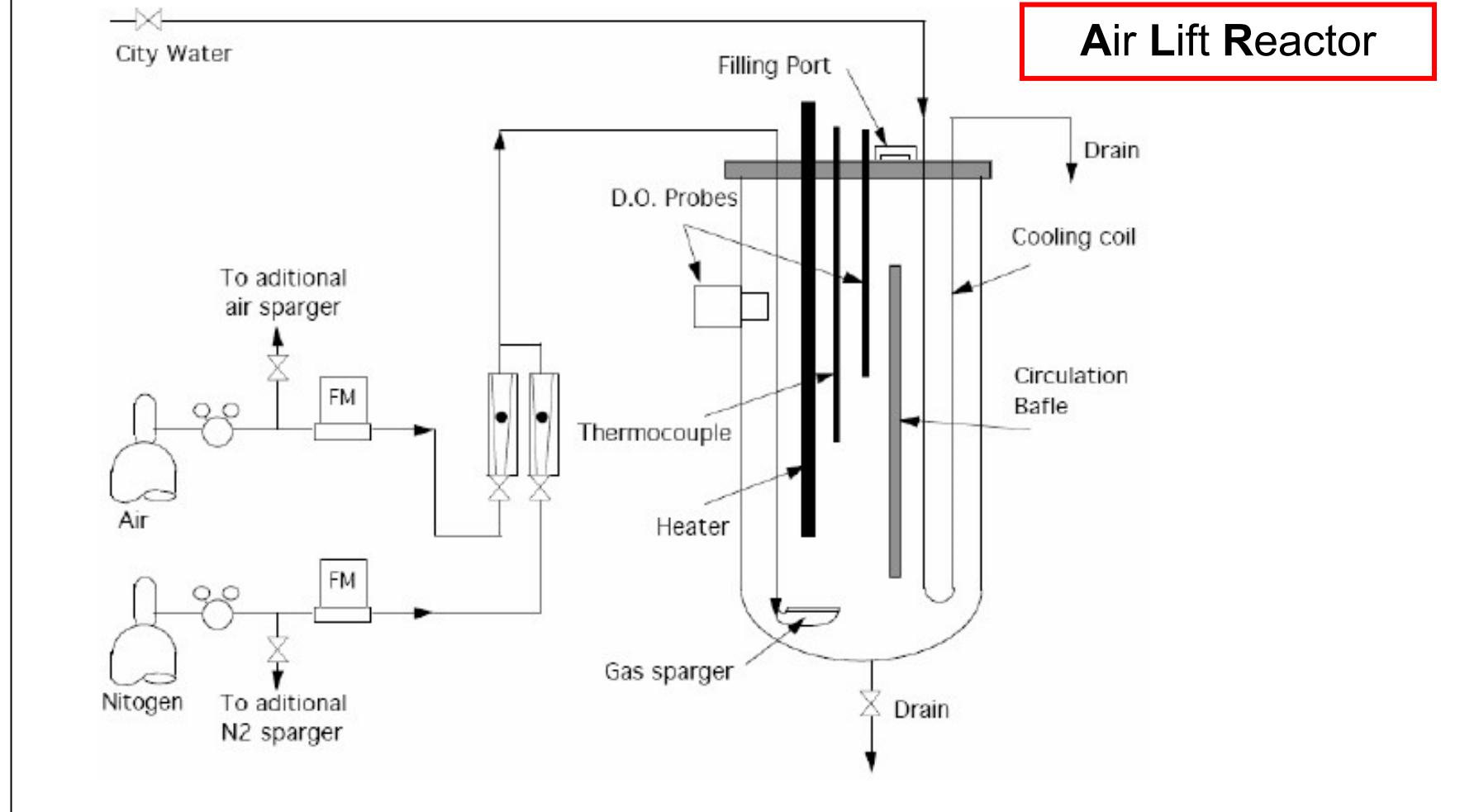
Loop reactors combining bubbles rising in a draft tube enhanced with  
a) stirring by an impeller  
b) Circulation of the liquid medium by a pump



Faculteit der Natuurwetenschappen, Universiteit van Amsterdam  
[http://www.science.uva.nl/research/cr/AirliftVib/experimental\\_setup.html](http://www.science.uva.nl/research/cr/AirliftVib/experimental_setup.html)

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# Submerged Fermentation

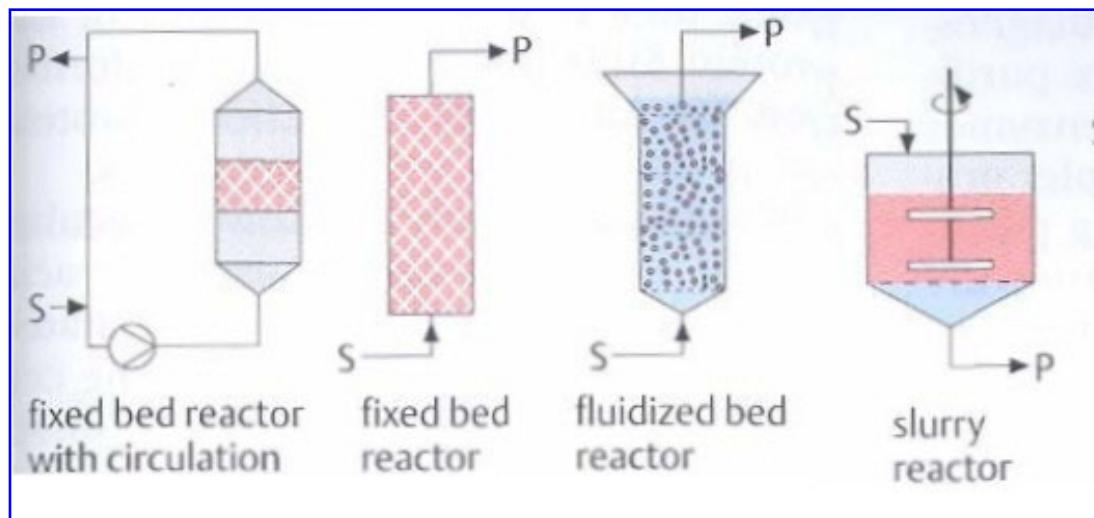
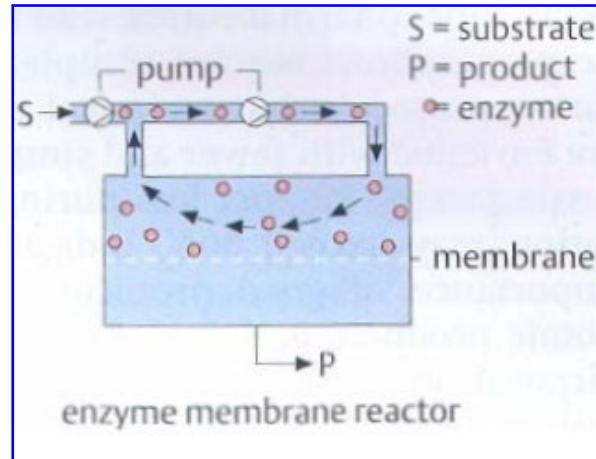


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# Submerged Fermentation

## others

- membrane bioreactors
- bed reactors

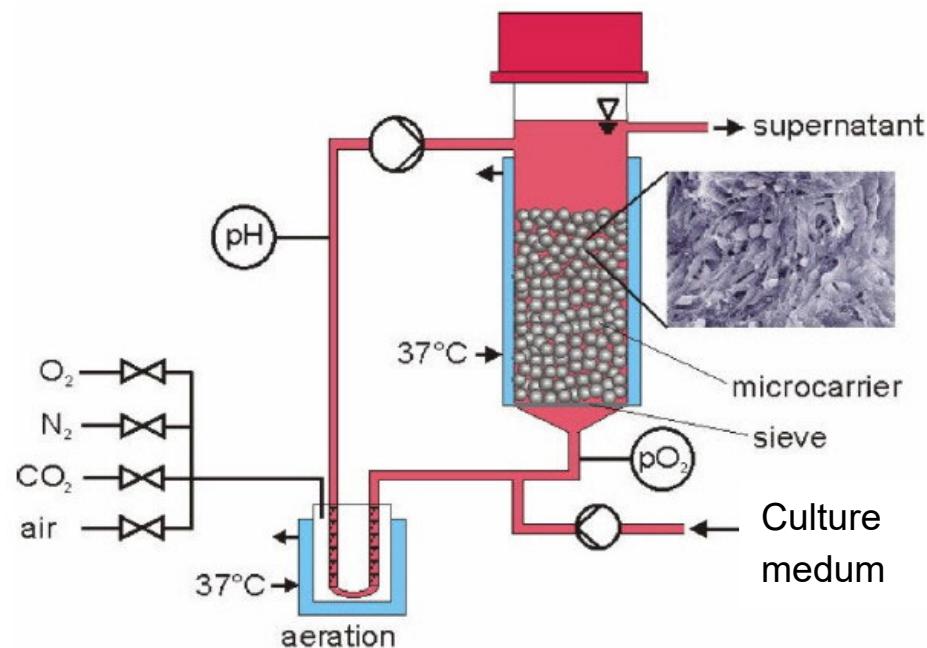


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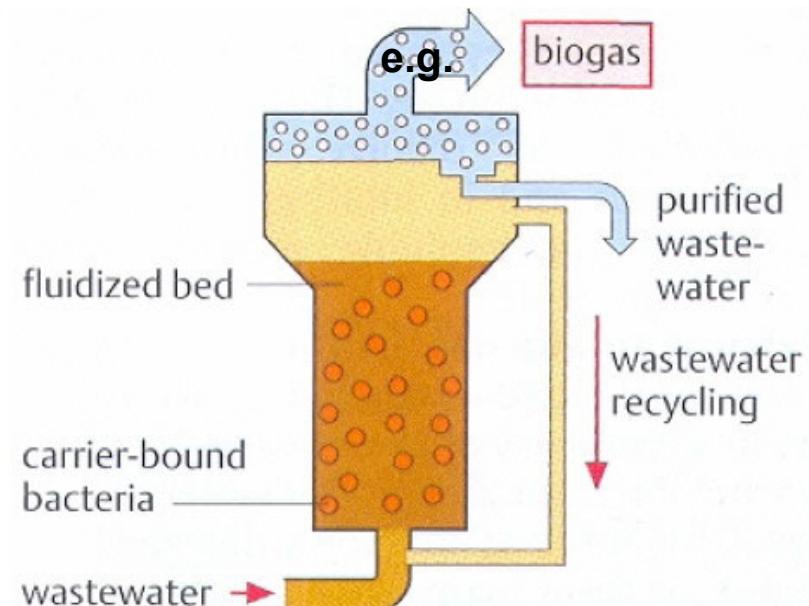
# Submerged Fermentation

others

- fixed bed reactors

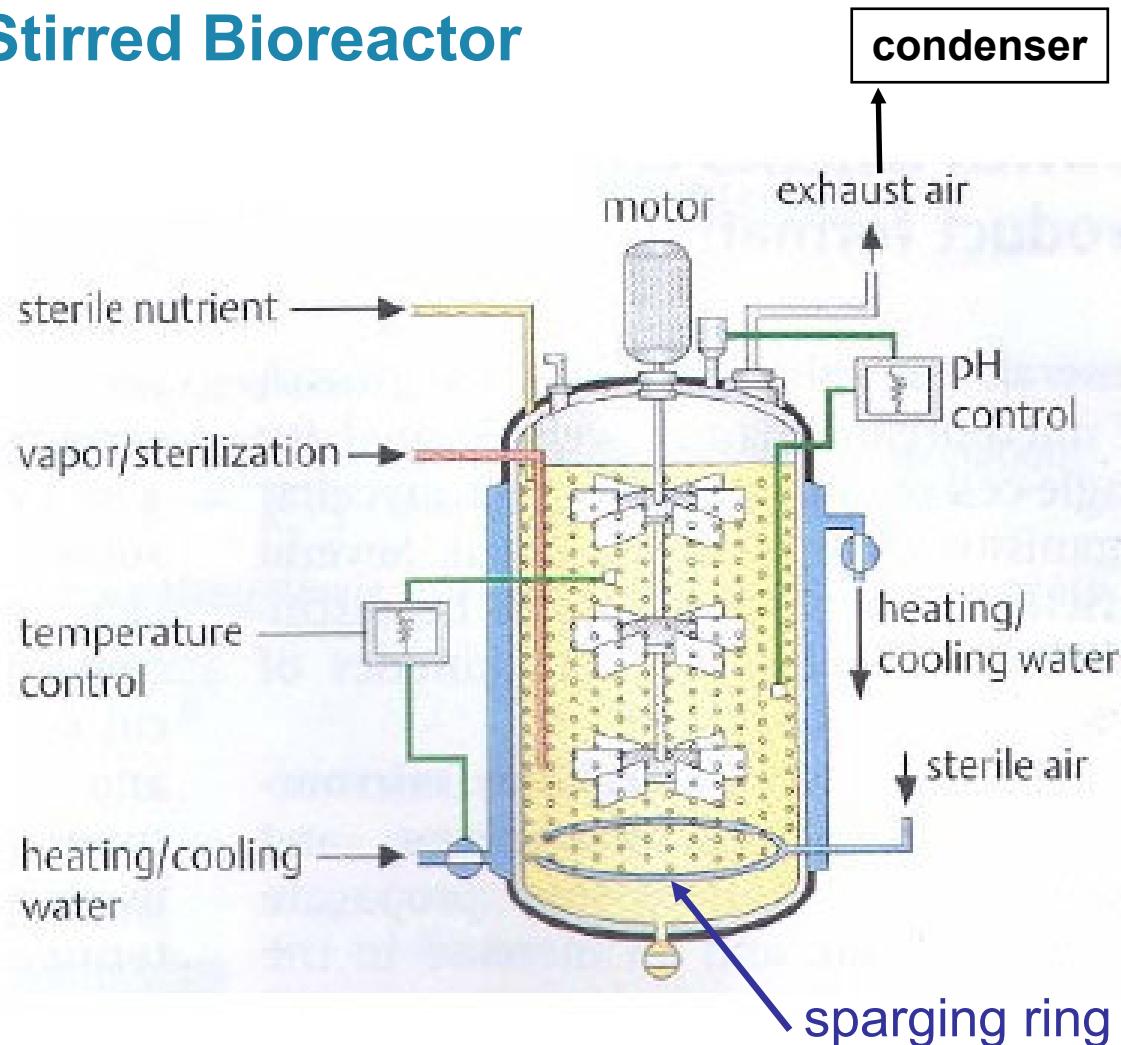


fluidized bed reactors



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# Stirred Bioreactor



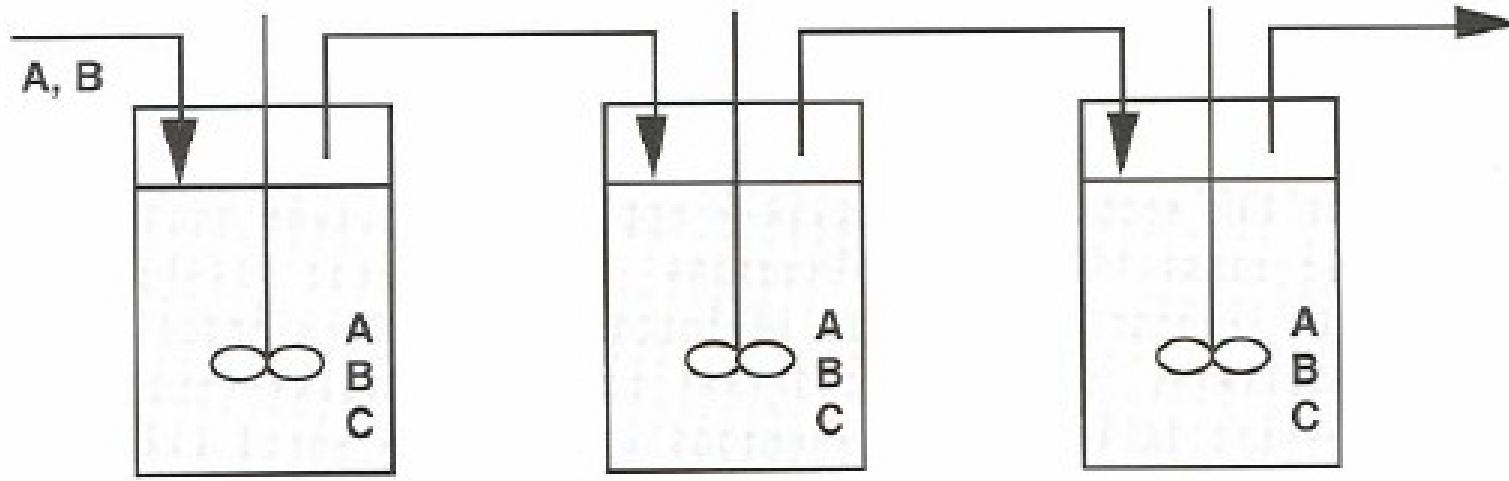
## Additional equipment

- feed addition port
- inoculum port
- sampling port
- pressure transducer
- agitator drive shaft bearing and seal
- foam breaker drive shaft bearing and seal

## Monitoring

- thermometer / temperature sensors
- pH electrode
- oxygen electrode
- possibly other electrodes/sensors
- foam sensor(s)

# Stirred-Tank Reactors in a Series



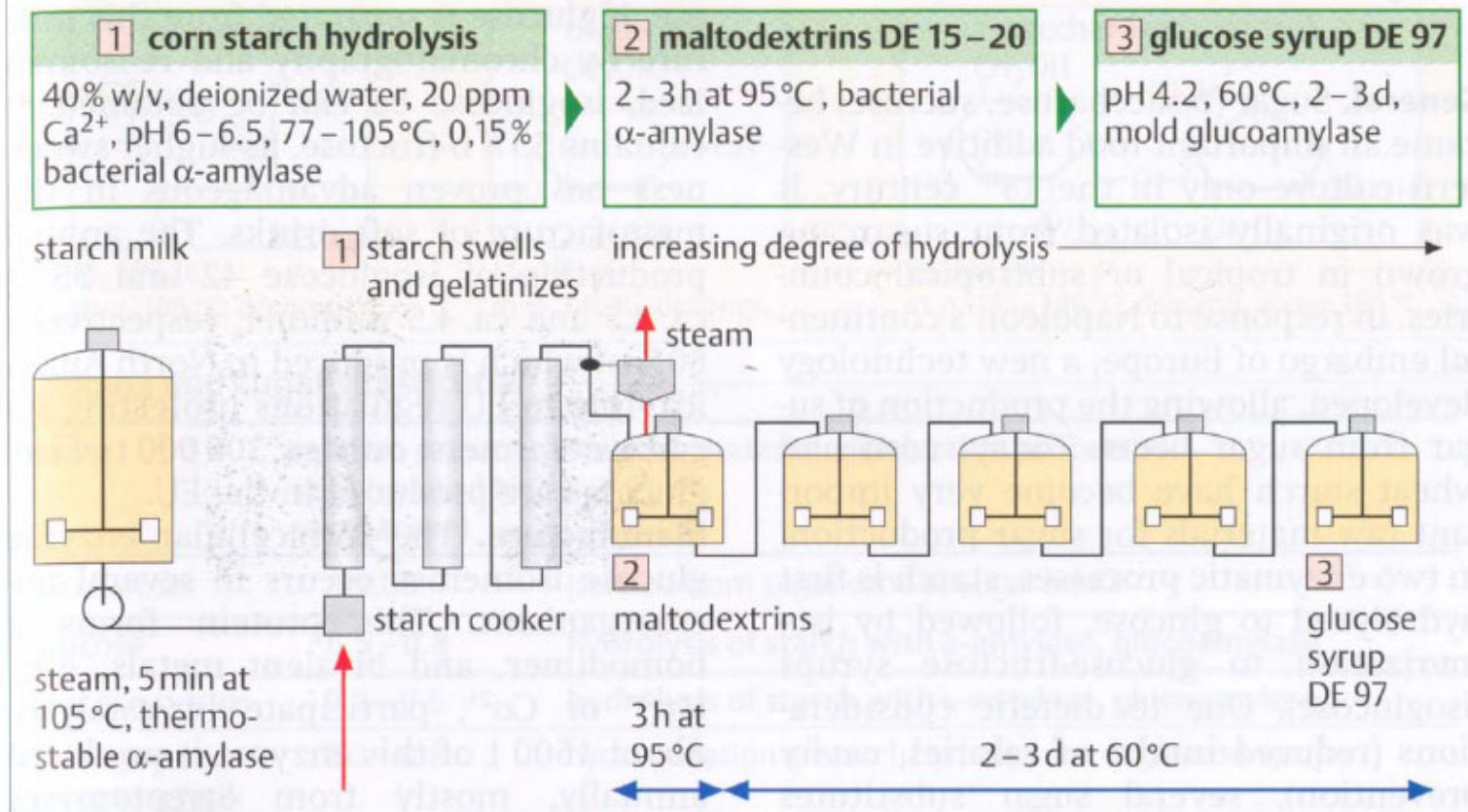
Stirred-tank reactors in series

- **cascade to produce the same product (C) in increasing concentrations (see figure)**  
or
- **production of consecutive products, e.g. hydrolyses of starch, a polysaccharide**

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# Enzymatischer Stärkeabbau in einer Rührkesselkaskade

## Enzymatic degradation of starch



R.D. Schmid; Pocket Guide to Biotechnology and Genetic Engineering, Wiley-VCH (2003); page 83

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## Batch reactor with a single external cooling jacket

Kühlmantel, besser Temperiermantel, *Heat transfer jacket* (bei Kultivierung von Säugerzellen dient der Mantel zum Temperieren auf 36-37 °C, beim Sterilisieren auf Temperaturen über 100 °C)



- outer jacket surrounding the vessel
- injection of heat transfer fluid at high velocity through nozzles, flow around the jacket
- Temperature in the vessel is regulated by changing the temperature of heat transfer fluid.

Bildquelle: Wikipedia, Creative Commons

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# Batch reactor with half coil jacket

Reaktor für Satzbetrieb mit Halbrohrschiene (zum Temperieren)



- made by welding a half pipe around the outside of the vessel
- → semi circular flow channel, heat transfer fluid passes in a plug-flow fashion
- for larger reactors: possibility to use several coils
- like in the case of the single external cooling jacket: regulation of vessel temperature by changing the temperature of heat transfer fluid

Bildquelle: Wikipedia, Creative Commons

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## Batch reactor with constant flux jacket (COFLUX)



A recent development

- series of >20 small jacket element
- temperature control by opening/closing valves to these jacket elements
- Temperature of the heat transfer fluid is kept constant.

Bildquelle: Wikipedia, Creative Commons

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# Measurement and Control in Bioreactor Technology

Physical parameters	Chemical parameters	Biological parameters
<ul style="list-style-type: none"><li>temperature</li><li>pressure</li><li>power input</li><li>viscosity</li><li>flow rate of air</li><li>nutrient feed</li><li>turbidity</li><li>bioreactor weight</li></ul>	<ul style="list-style-type: none"><li>pH value</li><li>dissolved O<sub>2</sub></li><li>O<sub>2</sub> and CO<sub>2</sub> in exhaust gas</li><li>redox potential</li><li>substrate concentration</li><li>product concentration</li><li>concentration of ions</li></ul>	<ul style="list-style-type: none"><li>enzyme activity</li><li>ATP content</li><li>NADH content</li><li>protein content</li></ul>

measured parameters

Deutsch: Messgrößen

regulated parameters  
set values  
Stellgrößen

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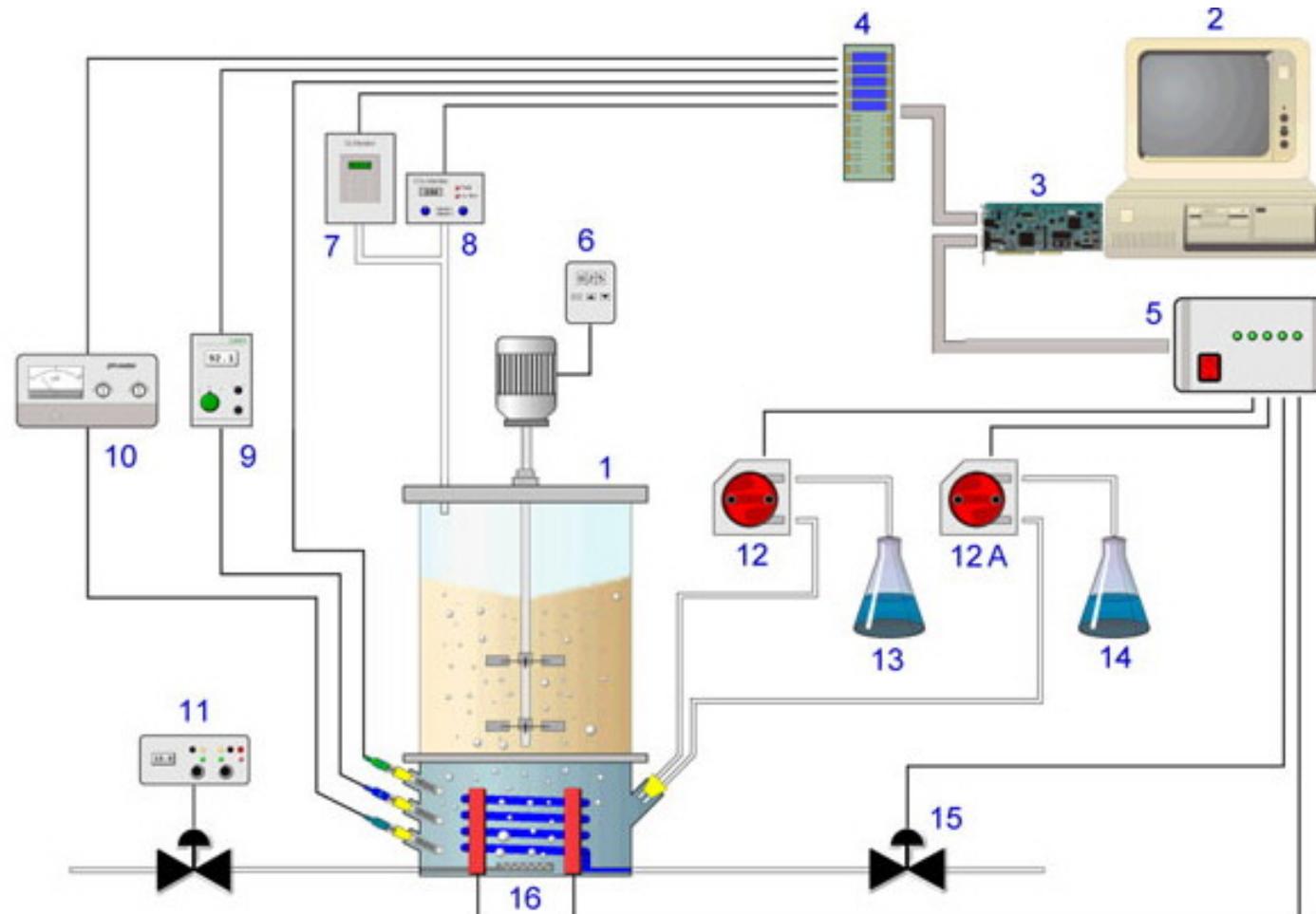
## On-line monitoring

Fermentations require constant control and adjustment:

- nutrient concentrations
  - bacteria concentrations
  - pH
  - temperature
  - contaminant concentrations (inlet and outlet)
  - vapour emissions
  - removal of solids
- **in case of aerobic fermentation:**
- oxygen levels & transfer efficiency

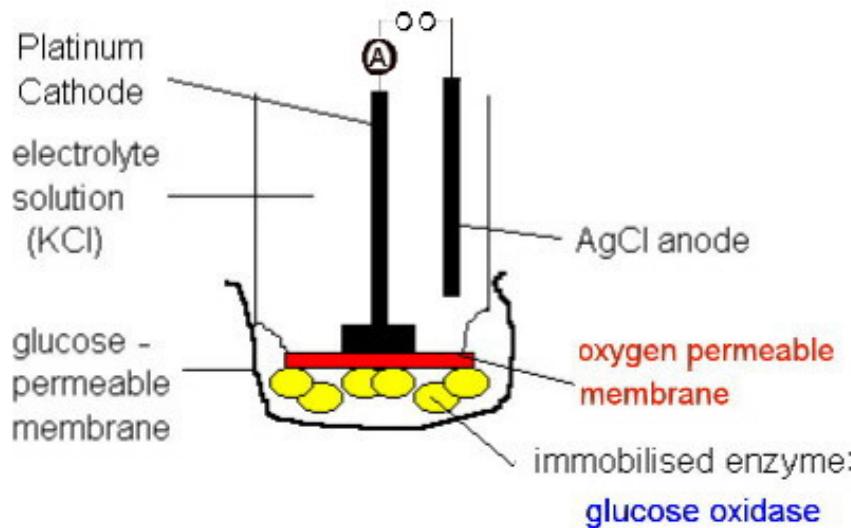
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# On-line monitoring



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# On-line monitoring of Substrate Concentration



Teeside University, [http://sst-web.tees.ac.uk/external/u0003076/BIOSENSORS\\_WEB/amperometric/clarketc.htm](http://sst-web.tees.ac.uk/external/u0003076/BIOSENSORS_WEB/amperometric/clarketc.htm), mod. Blokesch

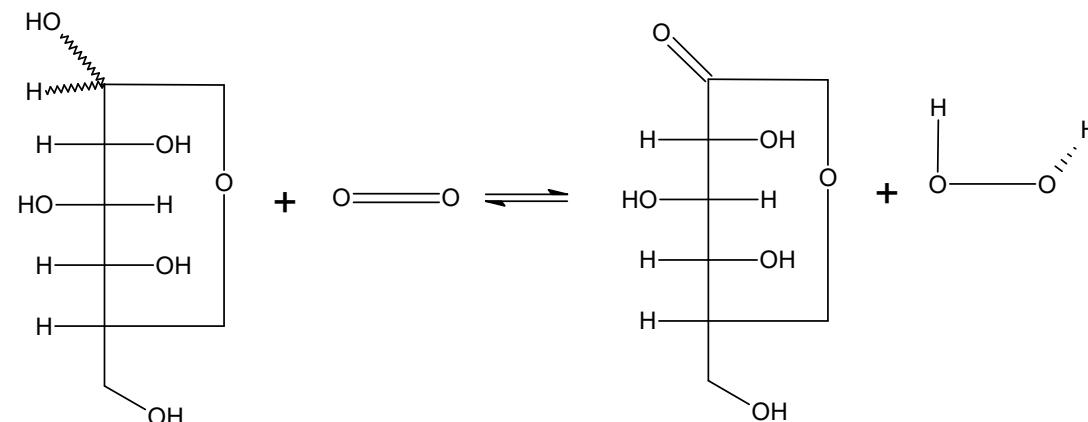
**example: Glucose Sensor by Amperometric Detection:**

either oxygen reduction at a Pt cathode

→ signal weakened in the presence of glucose

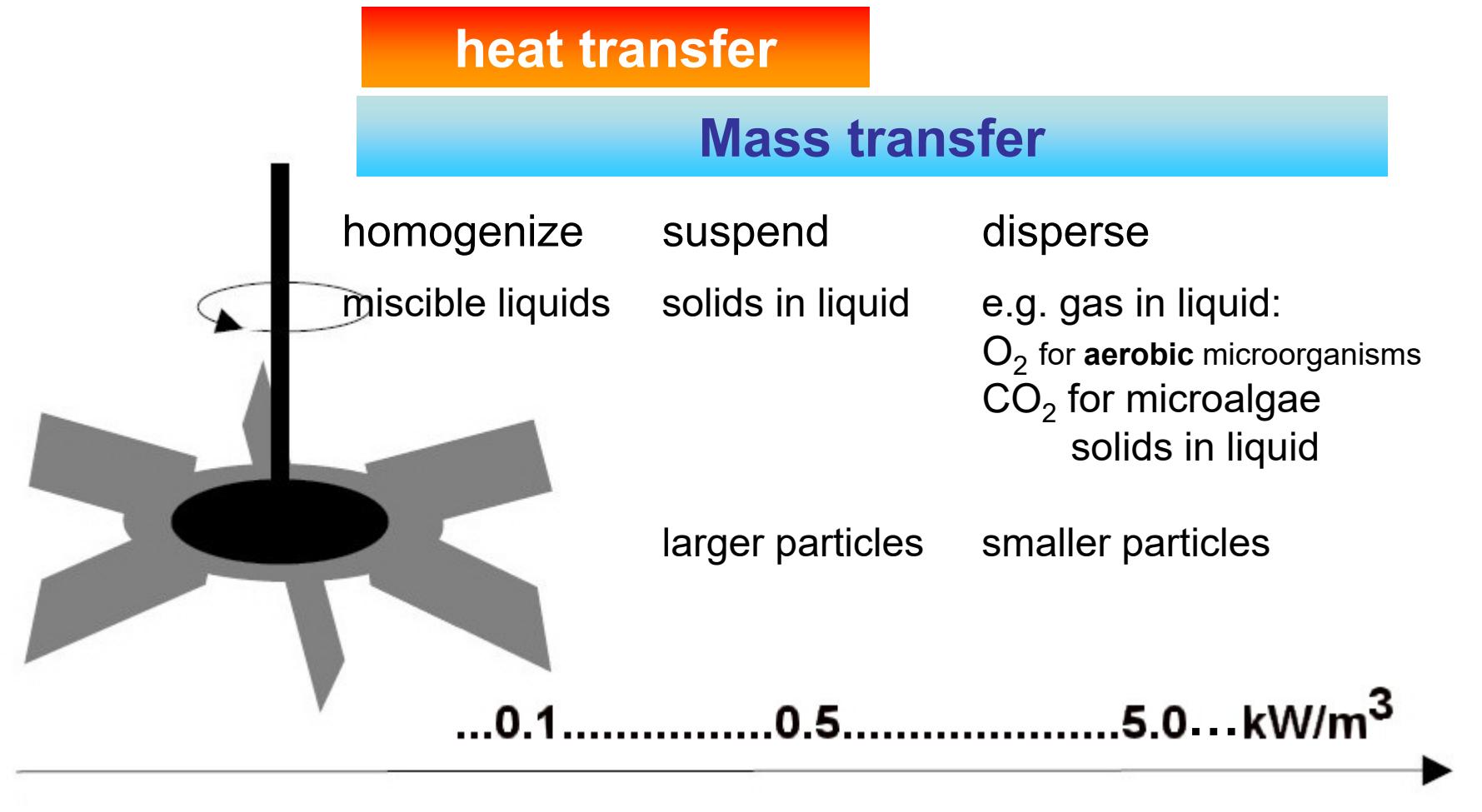
or  $\text{H}_2\text{O}_2$  oxydation to oxygen at a AgCl anode

→ signal enhanced in the presence of glucose

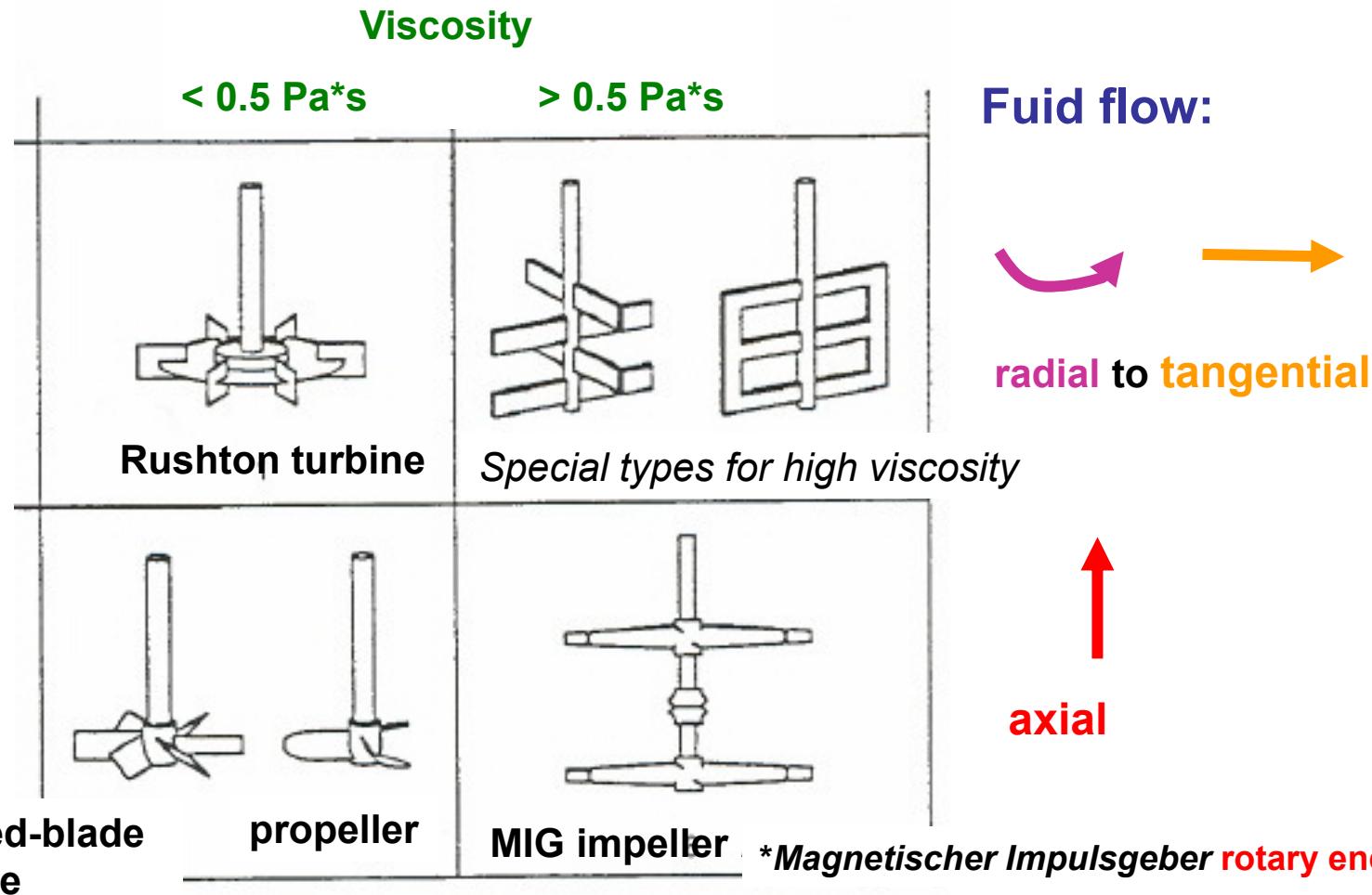


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# Purpose of Stirring

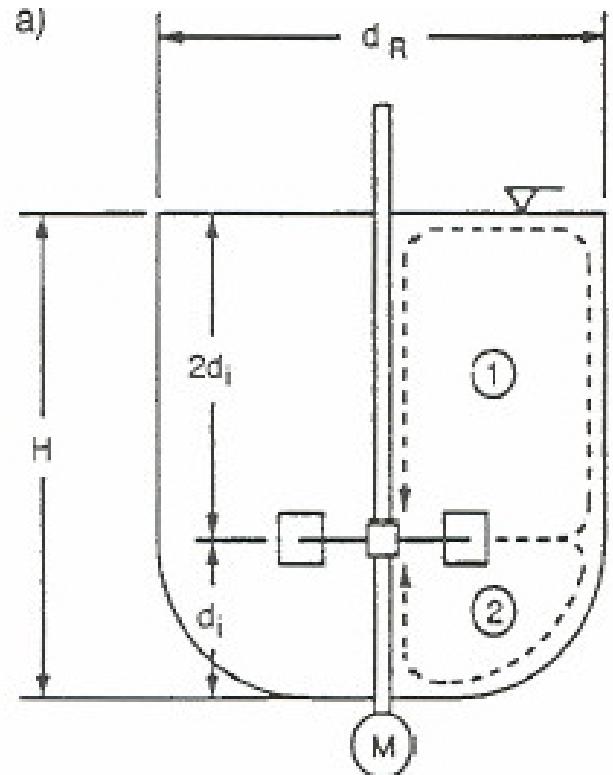


# Types of Common Impellers

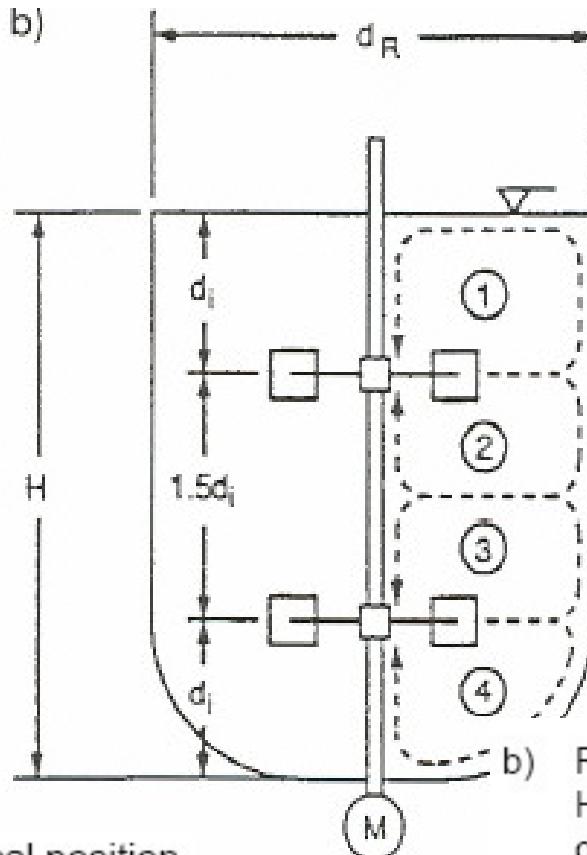


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## Flow pattern with circulation flow in stirred-tank reactors

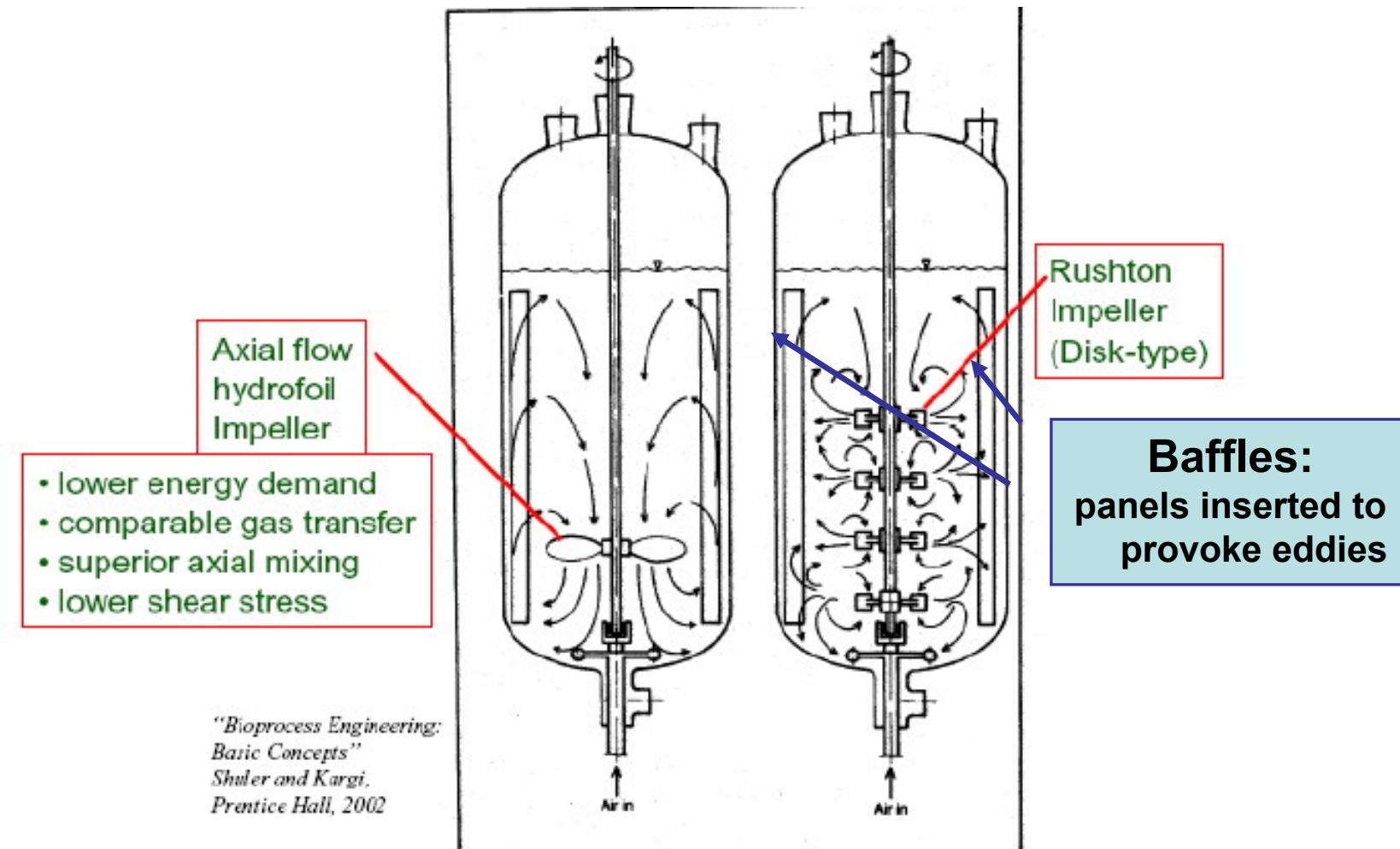


a) Reactor with impeller in unsymmetrical position



b) Reactor with two impellers  
 $H$  = height  
 $d_R$  = reactor diameter  
 $d_i$  = impeller diameter  
1 to 4 = circulation pathways

# Flow pattern with circulation flow in stirred-tank reactors



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# Power Transferred to the Medium by the Impeller

$$P_R = N_P \rho n^3 d_R^5$$

$P_R$  power released, i.e. transferred from impeller to medium

$N_P$  power number

in German: Ne Newton-Zahl, allgemein Verhältnis von Fließwiderstandszur Trägheitskraft einer Strömung, hier Anteil der Leistung des Rührwerks, die tatsächlich als hydraulische Leistung im Kessel zur Verfügung steht. Eine weitere wichtige Kennzahl ist die Reynolds Zahl Re, das Verhältnis von Trägheits- zu Zähigkeitskräften. Im Rührkessel ist bei  $Re > 10000$  der Übergang von laminarer zu turbulenter Strömung.

$\rho$  medium density

$n$  number of revolutions per time unit (usually minutes)

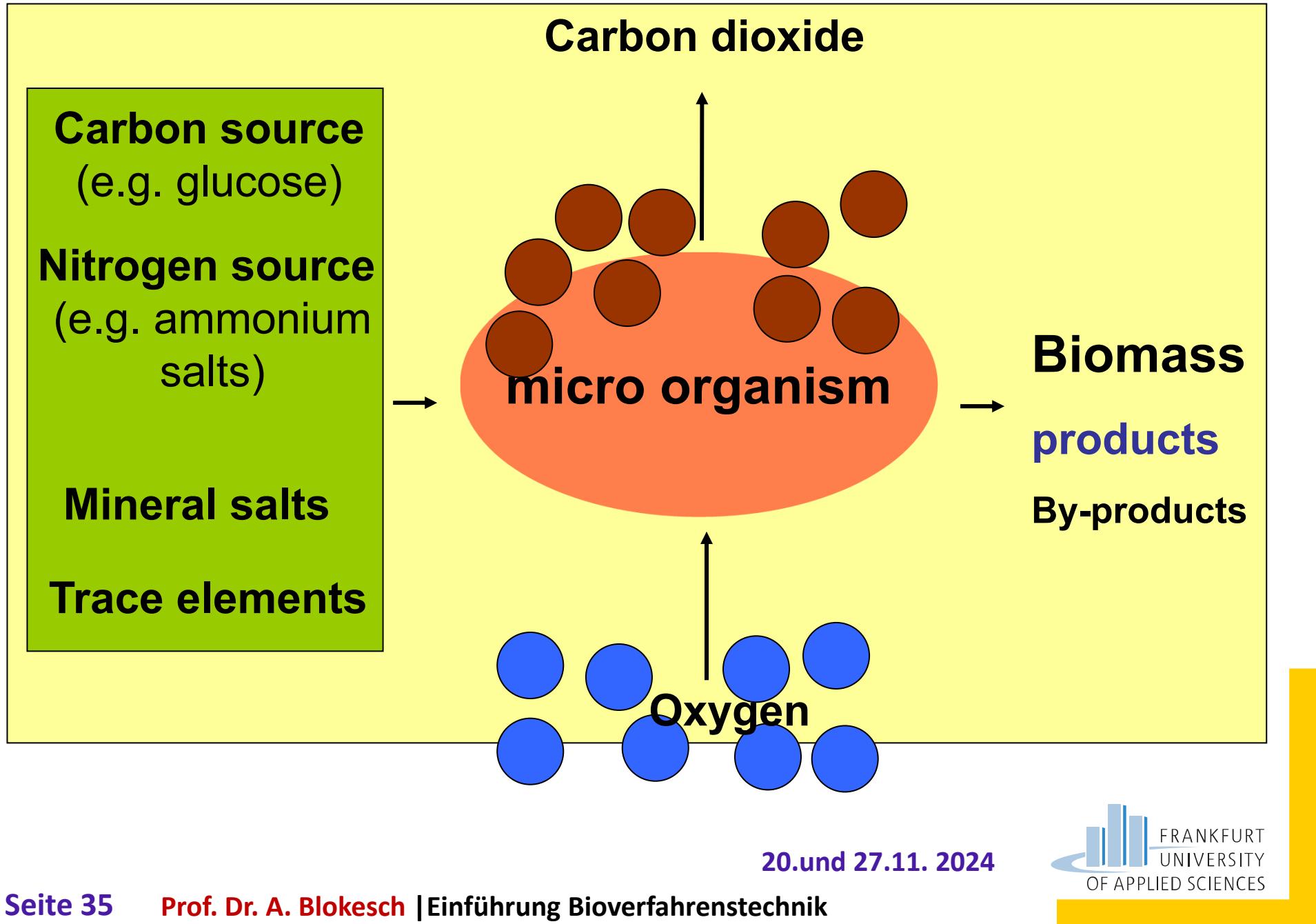
$d_R$  (inner) diameter of the rotor blades

## Neither stirring nor pressurized aeration needed:

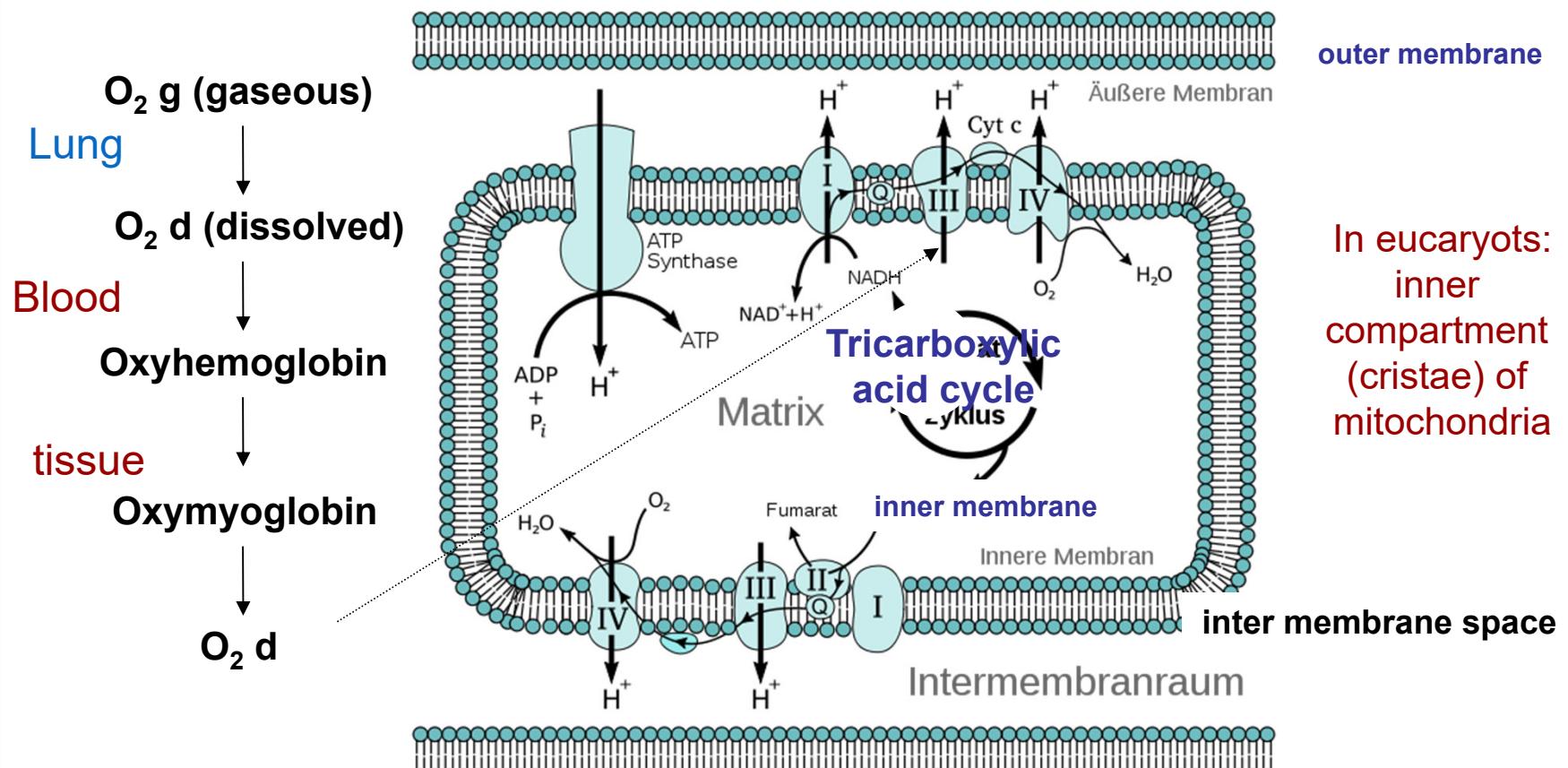
- traditional anaerobic fermentation processes in food production (e.g. beer, wine, dairy products)
- often large vats (= vessels)
- long-time fermentation, low growth rate

## Stirring and/or pressurized aeration:

- most industrial fermentations (aerobic) for the production of biotechnological products



# The Respiratory Chain



Source: [Klaus Hoffmeier](#), Übersetzung des engl. Originals von [Evasconcellos](#) 22:35, 9 September 2007 (UTC)

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## Critical Oxygen Concentration

Below that concentration, biomass growth and product formation of aerobically growing microorganisms decreases rapidly due to oxygen limitation.

Organism	$C_{crit}$ [mg/l]	$C_{crit}$ [ml/l]
<i>Escherichia coli</i>	0.26	0.20
<i>Penicillium chrysogenum</i>	0.40	0.30
<i>Saccharomyces cerevisiae</i>	0.60	0.46
<i>Pseudomonas ovalis</i>	1.10	0.84
<i>Torulopsis utilis</i>	2.00	1.52

Crueger, W., Crueger, A.: Biotechnologie – Lehrbuch der angewandten Mikrobiologie, Oldenbourg Verlag GmbH, Munich, Vienna, 1989.

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→ stoichiometric molar ratio

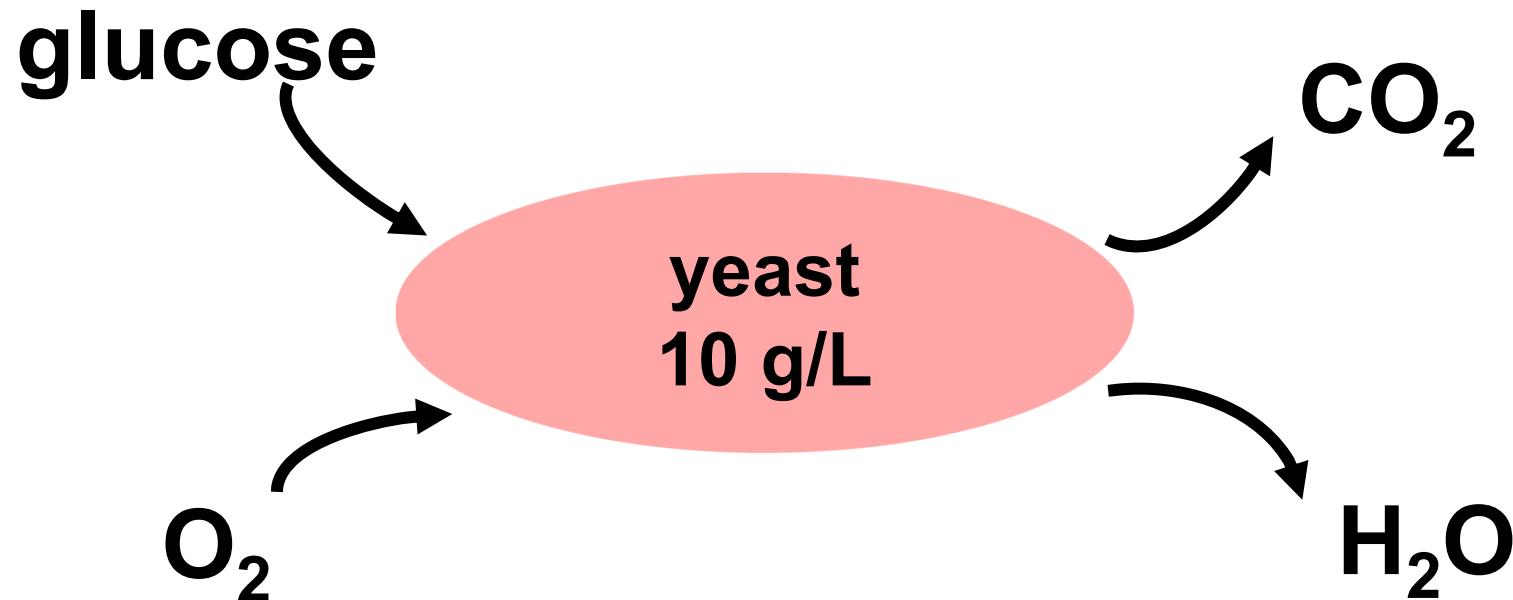
Glucose : Oxygen

1 : 6

real molar ratio in saturated solutions

1 :  $5 \cdot 10^{-5}$

## Model Fermentation – Necessity of Oxygen Supply



Oxygen demand:  $10 \text{ ml L}^{-1} \text{ min}^{-1}$ .

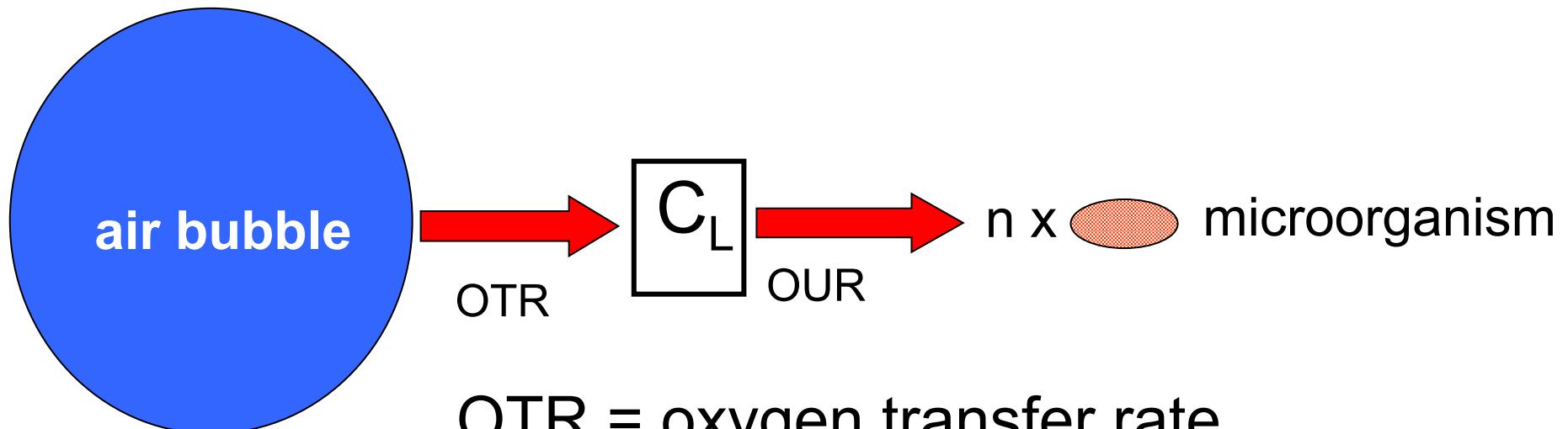
Maximal soluble concentration in medium:  $4 \text{ ml L}^{-1}$

Dissolved oxygen expires within **24 seconds**



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## Parameters Influencing O<sub>2</sub> Concentration in the Reactor



OTR = oxygen transfer rate

OUR = oxygen uptake rate

$$\frac{dC_L}{dt} = OTR - OUR$$

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## Oxygen Supply

Supplying sufficient oxygen represents one of the predominant and most challenging tasks for the operators of bioreactors.

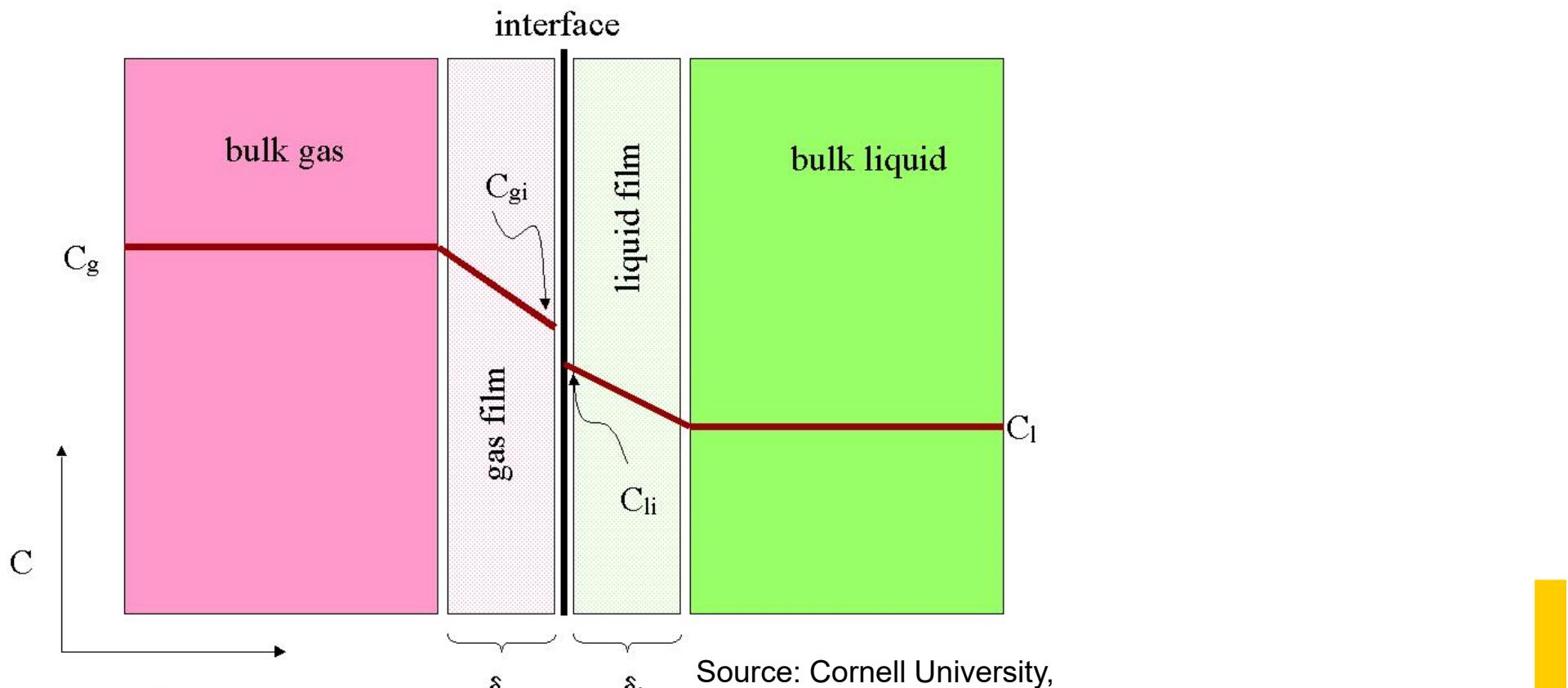
**OTR** = oxygen transfer rate  $\leq 3000 \text{ mL L}^{-1} \text{ h}^{-1}$   
gas → liquid medium

**OUR** = oxygen uptake rate  $\leq 4000 \text{ mL g}^{-1} \text{ h}^{-1}$   
liquid medium → microorganism

Please consider that biomasses formed during fermentation are often exceeding  $1 \text{ g L}^{-1}$ .

## Two film model

Developed Lewis-Whitman in 1923 – improving the previous film model proposed by Nernst in 1904 (with just one stagnant film at the interface)



Source: Cornell University,  
[ceeserver2.cee.cornell.edu/jjb2/cee6560/5-gas%20transfer.ppt](http://ceeserver2.cee.cornell.edu/jjb2/cee6560/5-gas%20transfer.ppt)

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## Penetration and Surface Renewal Models

More realistic models of the process have been proposed by Higbie (1935, penetration model) and by Danckwerts ( 1951, surface renewal model).

In these models bulk fluid packets (eddies) work their way to the interface from the bulk solution. While at the interface they attempt to equilibrate with the other phase under non-steady state conditions. No film concepts need be invoked. The concentration profile in each eddy ( packet) is determined by the molecular diffusion ...

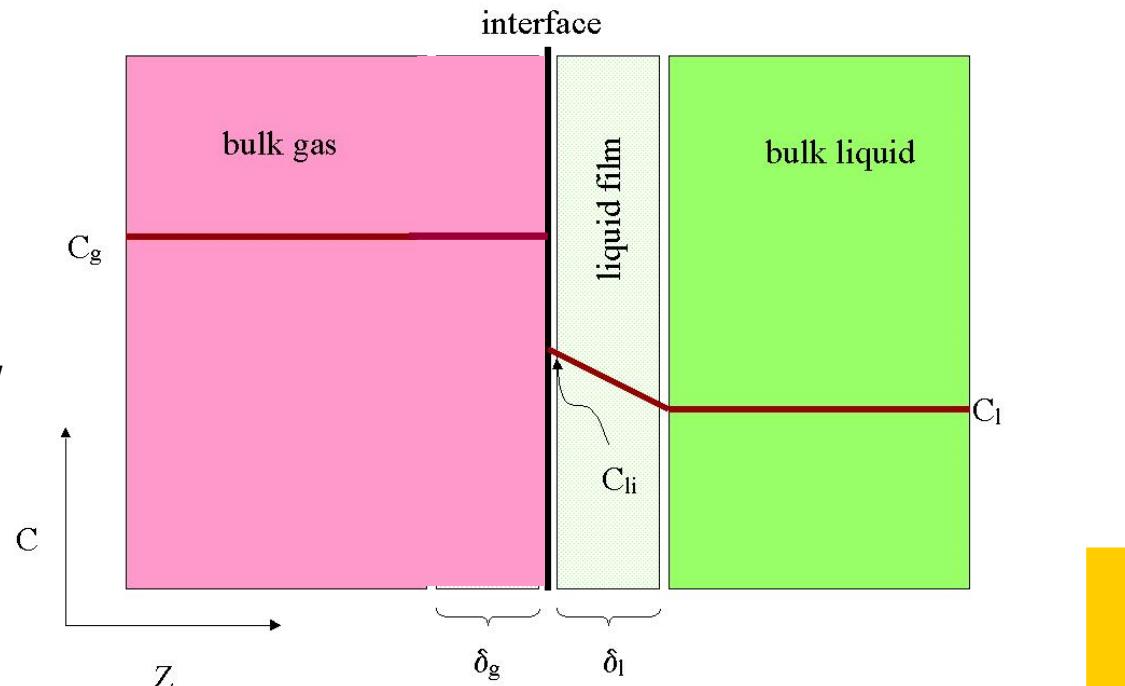
Source: Cornell University,  
[ceeserver2.cee.cornell.edu/jjb2/cee6560/5-gas%20transfer.ppt](http://ceeserver2.cee.cornell.edu/jjb2/cee6560/5-gas%20transfer.ppt)

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A simplified **Two film model** is used to explain oxygen transfer from the air bubble to the liquid medium because it is the easiest model for calculation.

Since oxygen transport within the gas film (*figure: left side*) is much faster than in the liquid film (*figure: right side*), that is  $k_g \gg k_l$ , only the transport in the liquid film is considered.

Figure: Cornell University,  
[ceeserver2.cee.cornell.edu/jjb2/cee6560/5-gas%20transfer.ppt](http://ceeserver2.cee.cornell.edu/jjb2/cee6560/5-gas%20transfer.ppt)



The time for oxygen transport from the liquid medium to the living cell is also neglected.

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The equilibrium of the partition of oxygen at the interface of gaseous and liquid phase is expressed by the law of Henry:

$$H * c_{li} = c_{gi}$$

Depends on:

- air pressure
- temperature
- salts and other compounds in the liquid media

	H	$c_{li}$
$p_{air} \uparrow$	=	$\uparrow$
$T \uparrow$	$\uparrow$	$\downarrow$
$c_{salt} \uparrow$	$\uparrow$	$\downarrow$

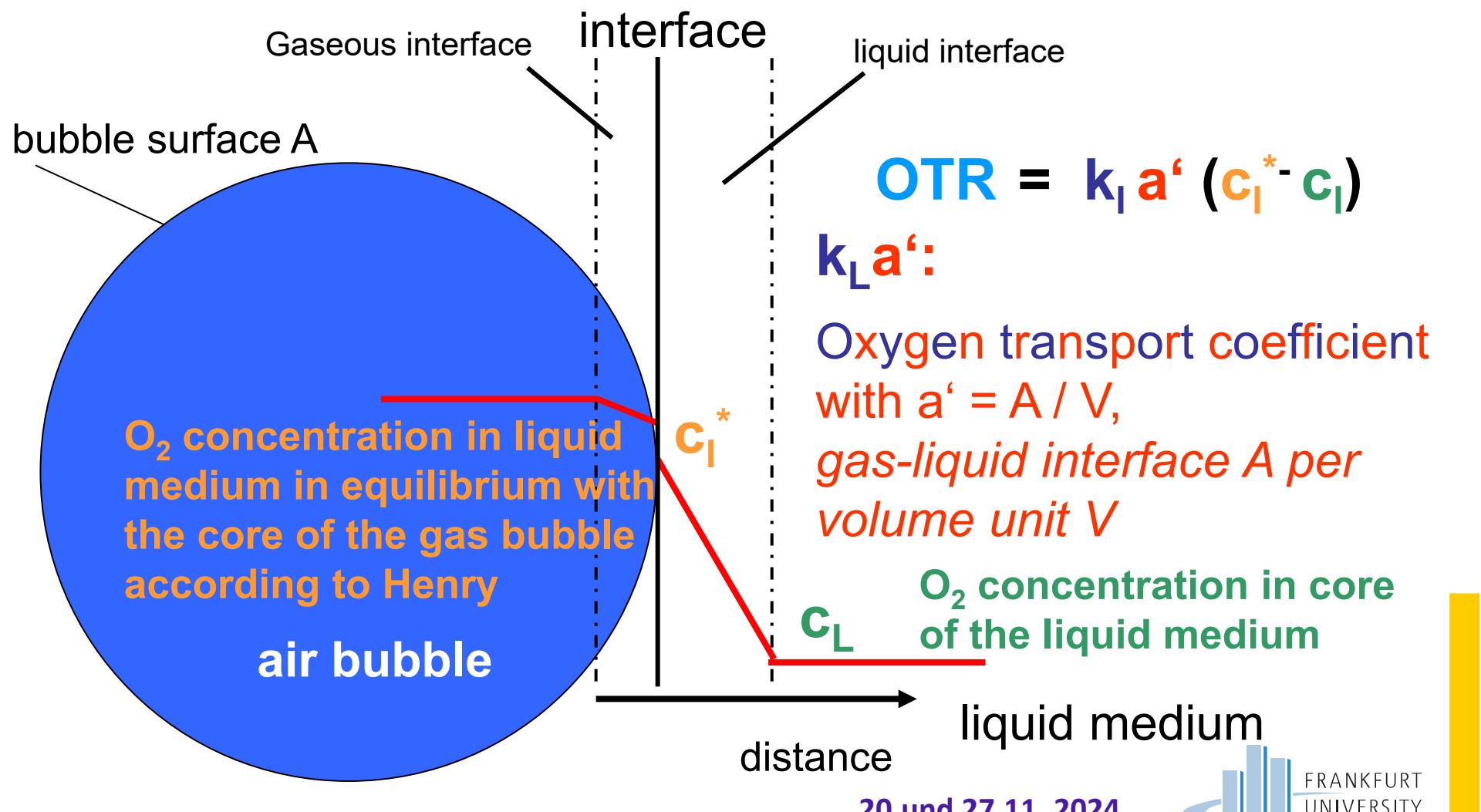
Oxygen Concentration in water at 20°C and 10<sup>5</sup> Pa air pressure, : 2\* 10<sup>4</sup> Pa partial pressure of O<sub>2</sub>:

0.275 mmol/l

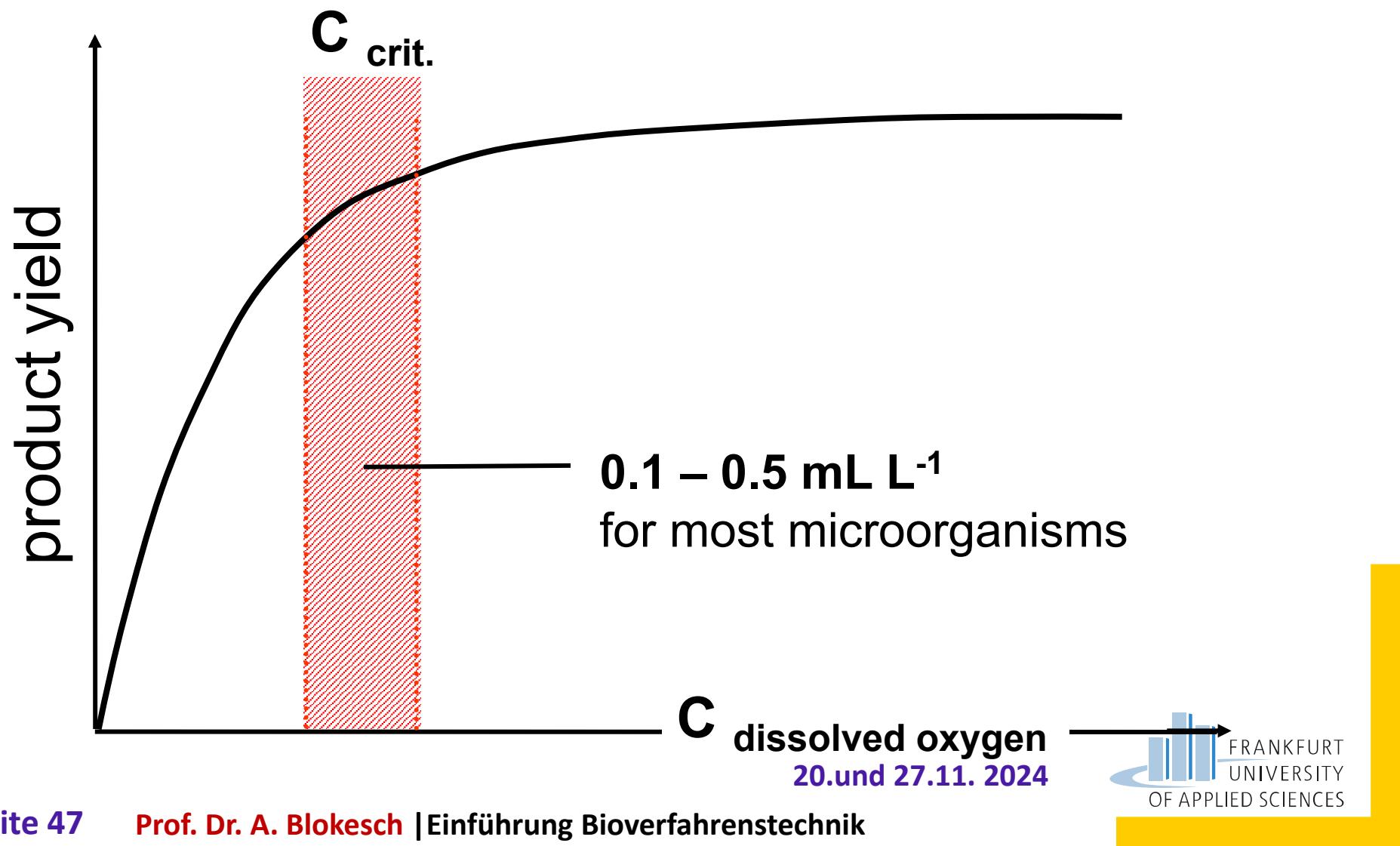
< 9 mg/l

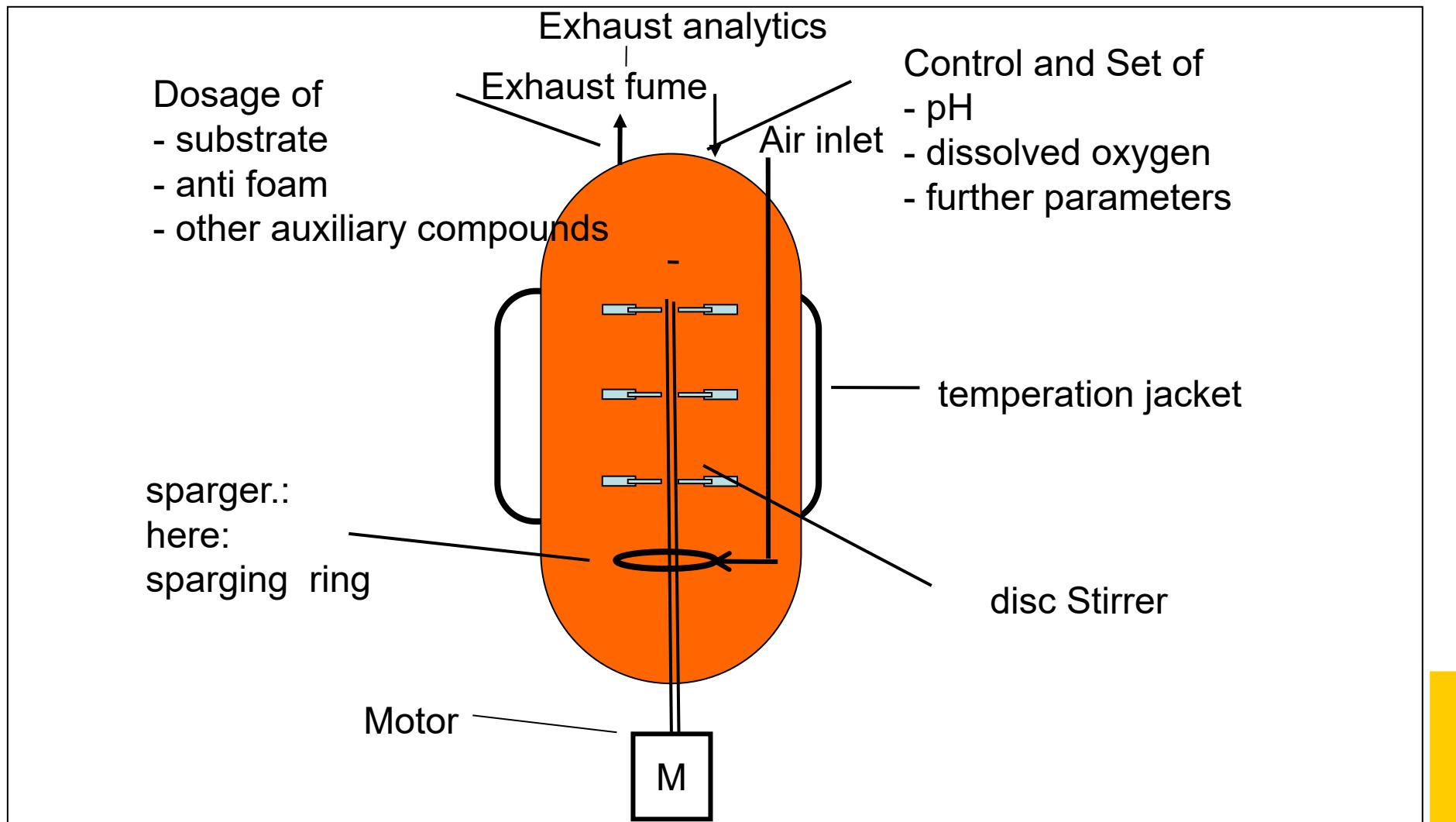
< 7 ml / l

## O<sub>2</sub> Concentration Gradient at the Interface of Gas to Liquid



## O<sub>2</sub> Concentration and Product Yield





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# Aeration

- sparger

- sparging frit



- sparging ring



Machine Engineering Faculty of the Dresden University of Technology (TU)

[http://mlu.mw.tu-](http://mlu.mw.tu-dresden.de/module/m018/Kap8/kap8_2/kap8_2_4/gasverteiler.html)

[dresden.de/module/m018/Kap8/kap8\\_2/kap8\\_2\\_4/gasverteiler.html](http://mlu.mw.tu-dresden.de/module/m018/Kap8/kap8_2/kap8_2_4/gasverteiler.html)

Accessed on 19-4-2010

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## Aeration

- sparger
- sparging frit
- sparging ring
- combined with stirring



Abb. 9 EKATO PHASEJET

gasjet propeller  
EKATO phasejet

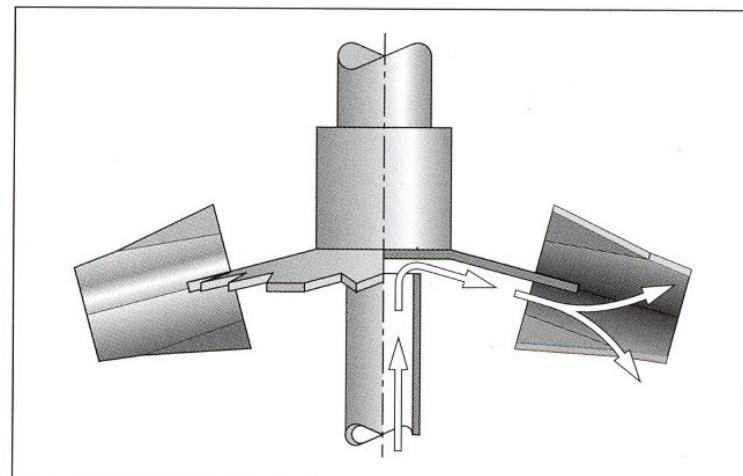


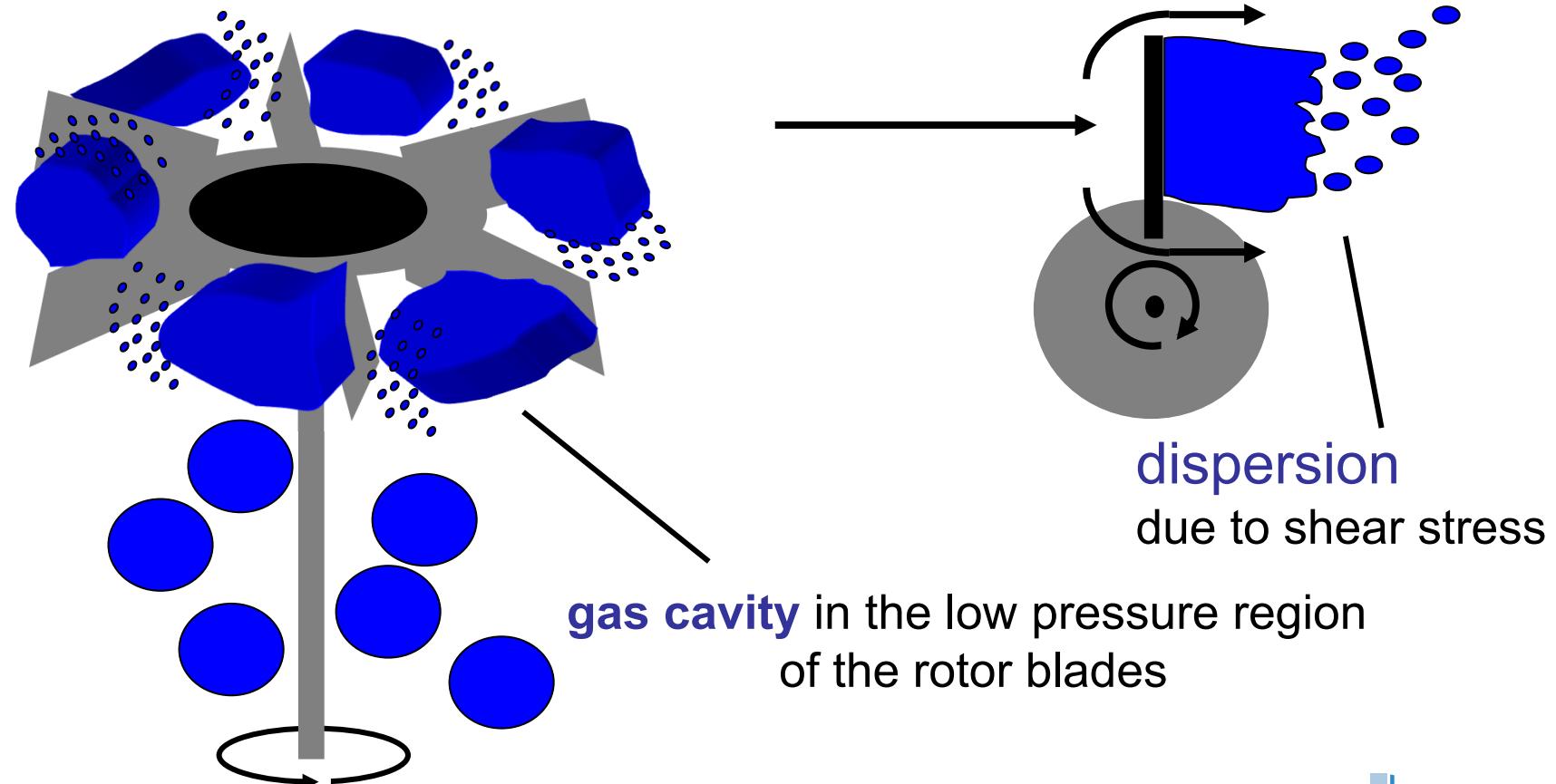
Abb. 10 Schnittbild des rotierenden Gasverteilers

rotating gas sparger

## Aeration

### Oxygen Transfer Rate (OTR)

Increased by Air Bubble Formation at the Impeller



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## Aeration – *an empirical formula*

$$k_L a \sim (P_{Rg} / V)^{0,7-0,8} V_s^{0,6}$$

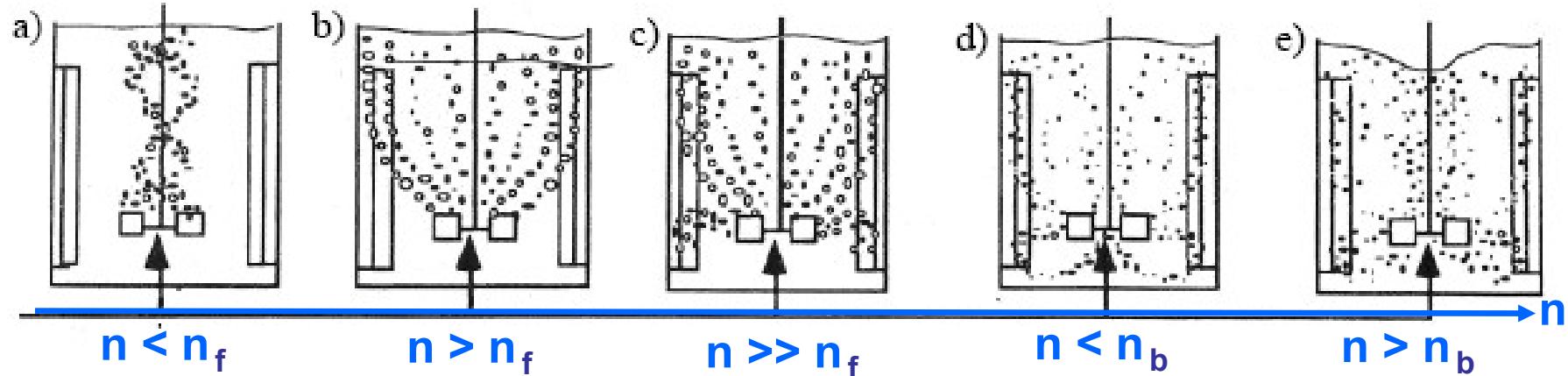
$P_{Rg}$  performance released by impeller in aerated state

$V$  volume of liquid in the reactor vessel

$V_s$  speed of gas flow in empty tube

## Gassing Modes

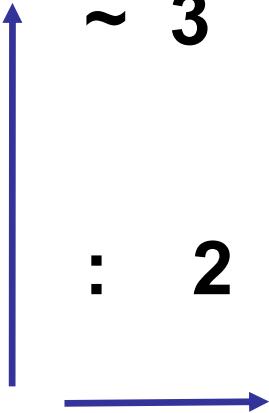
with a constant gas flow and increasing stirring speed  $n$



**Source:** Storhas, W.: Bioreaktoren und periphere Einrichtungen, Vieweg, Braunschweig, Wiesbaden 1994, transl. A. Blokesch

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# Reactor Geometry and Layout

- Height to diameter ratio of  $\sim 3$
  - sterile air inlet and sparger
  - impellers and baffle plates
  - cooling coils
  - foam breakers
  - working volume (liquid capacity)  $\leq 0.75$  vessel volume)
- 
- A vertical blue arrow points upwards from the text 'Height to diameter ratio of ~ 3'. To its right is a horizontal blue arrow pointing to the right, with the number '2' written vertically between them, indicating a 2:1 ratio.

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# Preparation of the Fermenter and Fermentation

## A) time of preparation

1. cleaning of the reactor
2. preparation of the substrate
3. sterilisation of the substrate (thermic or filter)
4. sterilisation of the reactor (cleaning in place)
5. calibration of the instruments of measurement (pH, pO<sub>2</sub>, pCO<sub>2</sub>, O<sub>2</sub>, CO<sub>2</sub>)
6. inoculation (monoculture or defined mixed culture)

## B) time of reaction (growth time)

7. fermentation (batch, fed-batch, repeated fed-Batch)
8. aeration
9. mixing (mechanical, pneumatical or hydrodynamical)
10. parameter control (with regulation, control, documentation)
11. sampling (manually or automatically)

## C) time of harvest

12. harvest
13. extraction of products
14. cleaning of the reactor

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## Explanations

**baffle** inserted panels close to the reactor vessel wall (in shaker flasks: notches of the glass wall into the inside space) conceived to disturb flow of fluid, provoke turbulence **Stromstörer**

**nozzle** narrow end of a tube (or hose), where fluid or gas jet issues (at high speed) **Sprühdüse**

**sparger** tube for moistening (= sprinkling), especially in brewery → **sparging ring, sparging frit**  
**Begasungsring, Begasungsfritte**

**eddie, vortex, whirlpool** fast circling motion of fluid or gas **Wirbel**

**plug-flow** flow in a tube in one direction and equal velocity along the tube diameter **Pfropfenströmung**

**hamster** small rodent (same term as in German)